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A Bayesian non-linear mixed effects regression model for the characterisation of early bactericidal activity of tuberculosis drugs

Technical report prepared for Global Alliance for TB Drug Development (TB Alliance)

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ABSTRACT

Trials of the early bactericidal activity (EBA) of tuberculosis (TB) treatments assess the decline, during the first few days to weeks of treatment, in colony forming unit (CFU) count of *Mycobacterium tuberculosis* in the sputum of patients with smear-microscopy-positive pulmonary TB. Profiles over time of CFU data have conventionally been modelled using linear, bi-linear or bi-exponential regression. We propose a new bi-phasic non-linear regression model for CFU data that comprises linear and bi-linear regression models as special cases, and is more flexible than bi-exponential regression models. A Bayesian non-linear mixed effects (NLME) regression model is fitted jointly to the data of all patients from a trial, and statistical inference about the mean EBA of TB treatments is based on the Bayesian NLME regression model. The posterior predictive distribution of relevant slope parameters of the Bayesian NLME regression model provides insight into the nature of the EBA of TB treatments; specifically, the posterior predictive distribution allows one to judge whether treatments are associated with mono-linear or bi-linear decline of log(CFU) count, and whether CFU count initially decreases fast, followed by a slower rate of decrease, or *vice versa*.

1 INTRODUCTION

1.1 Early Development of Tuberculosis Treatment Regimens

Standard efficacy endpoints in pivotal Phase III trials of tuberculosis (TB) treatments are the proportion of patients with positive sputum culture after 6 months of treatment, and the proportion of patients experiencing relapse within a two-year follow-up period (Mitchison, 2006; Mitchison and Davies, 2008). Proof of clinical efficacy of TB treatments, therefore, generally requires lengthy and expensive clinical trials (Mitchison, 2006; Phillips and Fielding, 2008; Wallis et al., 2009). Furthermore, mono-therapy with anti-TB drugs is often ineffective, mainly due to increasing incidence of drug-resistance (Yang et al., 2011), so that TB is typically treated with combinations of bactericidal and sterilising drugs (Diacon et al., 2012a). As Diacon et al. (2012a) state, "ideally [new treatment] regimens would contain new drugs able to combat tuberculosis resistant to currently available drugs, especially multidrug-resistant (MDR) tuberculosis ...". Thus one of the challenges in early development of new TB treatments is to identify promising combinations of drugs for subsequent testing in pivotal clinical trials. Since the treatment regimens may involve combinations of three or four drugs, including one or more novel molecules, potentially large numbers of regimens need to be screened. One way to do so efficiently and cost effectively is to assess the early bactericidal activity (EBA) of those regimens.

1.2 Early Bactericidal Activity

An EBA trial assesses the decline, during the first few days to weeks of treatment, in colony forming unit (CFU) count of *Mycobacterium tuberculosis* in the sputum of patients with smear-microscopy-positive pulmonary TB (Diacon et al., 2012a). Such EBA trials

are usually conducted during the early stage of drug development (Phase II).

An early definition of EBA was the "fall in counts/mL sputum/day [of CFU count] during the first two days of treatment" (Mitchison and Sturm (1997) as cited in Donald and Diacon (2008)). More generally, the EBA in a given patient over a time interval from Day t_1 to Day t_2, i.e. EBA$(t_1 - t_2)$, can be estimated as follows:

$$\text{EBA}(t_1 - t_2) = -\frac{\log(\text{CFU}_{t_2}) - \log(\text{CFU}_{t_1})}{t_2 - t_1} \tag{1}$$

(see, e.g. Botha et al. (1996)). Here, $\log(\text{CFU}_{t_1})$ and $\log(\text{CFU}_{t_2})$ are the observed $\log(\text{CFU})$ counts at Day t_1 and Day t_2, respectively, where $0 \leq t_1 < t_2 \leq T$, and T is the length of the profile period over which serial sputum samples are collected. Equation (1) represents a "model-free" estimate of EBA$(t_1 - t_2)$, since it is function only of the observed $\log(\text{CFU})$ counts at Day t_1 and Day t_2.

Alternatively, EBA$(t_1 - t_2)$ can be estimated as:

$$\text{EBA}(t_1 - t_2) = -\frac{\hat{f}(t_2) - \hat{f}(t_1)}{t_2 - t_1} \tag{2}$$

where $f(t)$ is a suitable regression function for $\log(\text{CFU})$ count versus time, and $\hat{f}(t_1)$ and $\hat{f}(t_2)$ are the associated fitted values at Day t_1 and Day t_2, respectively (see, e.g. Jindani et al. (2003)).

The model-based estimate of EBA$(t_1 - t_2)$ in Equation (2) has two potential advantages over the model-free estimate in Equation (1): Firstly, the EBA estimate in Equation (1) uses information from only two CFU counts, namely those observed at Day t_1 and Day t_2; in contrast, the whole series of observed CFU counts may be used to estimate $f(t_1)$ and $f(t_2)$, with potential gains in precision for the model-based EBA estimate in Equation (2). Secondly, the model-free EBA estimate for a given time interval $(t_1 - t_2)$ can only be calculated if CFU counts are in fact available for these particular

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times; in contrast, the model-based estimate can be calculated (e.g. by extrapolating the curve over time interval $(t_1 - t_2)$) even if CFU counts have not been observed at Day t_1 and Day t_2, either because the study design did not specify data collection at those times, or because of missing data.

We note that, if the regression function $f(t)$ is linear over the whole profile period $[0, T]$, then the EBA estimate in Equation (2) is given by minus one times the slope of the regression of log(CFU) count against time. Indeed, both *in vitro* and *in vivo* studies have suggested that anti-TB drugs eradicate a fixed proportion of TB bacteria per unit time (Gillespie et al., 2002), at least over suitably short time intervals, which would imply an exponential decay in CFU count, or equivalently, a linear decay in log(CFU) count. Thus, if the decay of CFU counts over the *whole* interval $[0, T]$ is exponential (equivalently, log-linear), the EBA estimate in Equation (2) over all sub-intervals $(t_1 - t_2)$ of $[0, T]$ is constant, and equal to minus one times the slope of the linear regression line of log(CFU) count versus time.

1.3 Need for Non-Linear Regression Models

Jindani et al. (2003) argued that "standard EBA" TB trials, namely those estimating EBA(0-2), may fail to measure the sterilising activity of TB drugs: For example, monotherapy of pyrazinamide has been shown to be less bactericidal than that of isoniazid and streptomycin during the first few days of treatment (EBA), but proves to eradicate TB bacteria about the same rate afterwards (sterilisation). Thus, even though pyrazinamide has weak EBA, its sterilising activity proves to be better than that of isoniazid and streptomycin (Brindle et al., 2001; O'Brien, 2002). Based on these findings, Jindani et al. (2003) suggested the extension of "standard EBA" trials to a treatment period of at least 5 to 7 days, in order to evaluate the sterilisation activity of anti-TB drugs. Currently, the treatment and profile period for EBA trials typically is 14 days, with collection of one

or two pre-treatment and serial post-treatment overnight sputum samples. EBA values that are routinely reported for such TB trials include EBA(0-2), EBA(0-14), EBA(2-14), EBA(7-14).

As mentioned above, over a suitably short time interval a TB drug typically eradicates a fixed proportion of TB bacteria per unit time, implying exponential decline of CFU count over the time interval in question. Empirically, an exponential decline of CFU counts (or a linear decline in log(CFU) counts) has indeed been observed for most TB regimens, at least during the first few days of treatment, and certainly during the first two days. Thus, EBA(0-2) can be estimated from a simple linear regression of log(CFU) count versus time (see Equation (2)) (Brindle et al., 2001; Jindani et al., 2003; Dietze et al., 2008). However, when the profile period of EBA trials, and associated EBA calculations, covers time intervals significantly longer than 2 days, say 14 days, then the assumption of a constant rate of decay over the whole time interval generally is no longer valid. In fact, for many TB drugs, a significant difference between the rate of decline over the first two days of treatment compared to the subsequent days has been observed (Donald and Diacon, 2008): Usually, during the first few days of treatment, log(CFU) counts decline with a fast rate, followed by a slower rate of decline during the second phase. The decline in log(CFU) count can therefore be bi-phasic (Mitchison and Davies, 2008) over a 14-day treatment period. Thus, for EBA trials with longer profile periods, estimation of EBA generally requires some form of non-linear modelling that appropriately reflects the bi-phasic nature of the regression of log(CFU) count against time.

1.4 Non-Linear Regression Models Proposed in Literature

In order to account for the bi-phasic nature of log(CFU) count versus time curves, two types of non-linear regression models have essentially been described in the literature,

namely bi-linear and bi-exponential regression.

Diacon et al. (2012a, 2013) performed bi-linear regression of log(CFU) count against time on a by-patient basis, with visual identification of the node parameter (or inflection point), and assuming that the node was the same for all patients in a given treatment group. Thus, the approach of Diacon et al. (2012a, 2013) did not accommodate between-patient variation in the node. Accordingly, EBA was compared between treatment groups using analysis of variance (ANOVA) of the resulting by-patient EBA estimates. Furthermore, it would seem preferable to estimate the node parameter from the data, rather than determine it through visual inspection. In addition, it would seem preferable to fit the model as a bi-linear mixed effects regression model.

Jindani et al. (2003) suggested that the switch of one rate of decline in log(CFU) count to another might be smooth (rather than abrupt, as would be implied with a bi-linear regression model). Modelling such a smooth transition, Gillespie et al. (2002) and Jindani et al. (2003) used bi-exponential regression of CFU count against time, while Davies et al. (2006a), Davies et al. (2006b) and Rustomjee et al. (2008) regressed log(CFU) count, observed over 56 days of treatment, against the logarithm of a bi-exponential function as a mixed effects regression model. However, in bi-exponential regression models, the initial rate of decline in CFU count necessarily is greater than the terminal rate. Thus, bi-exponential regression models do not seem adequate for treatments (and individual profiles) which are associated with terminal rates of decline that are faster than initial rates of decline. Such treatments have only been described recently (Diacon et al., 2012a). The bi-exponential mixed effects regression model can fit data beyond 14 days of treatment, e.g. for 56-day "serial sputum colony counts (SSCC)" trials (Rustomjee et al., 2008). The trial discussed by Rustomjee et al. (2008) shows a clear distinction between the EBA and longer term sterilising activity for each of the treatment regimens: More specifically, per treatment group, the mean log(CFU) count

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over time suggests that the initial slope is substantially larger than the terminal slope. In our experience, the attempt to fit such a model to data beyond the scope of 14-day EBA trials results in convergence issues when the terminal slopes are greater than the initial slopes.

1.5   Objectives and Outline of Present Paper

The observations in the above section indicate that non-linear regression models for log(CFU) count versus time data published in the literature might require some modification and generalisation. In this paper, we propose a new non-linear regression model for log(CFU) count (Burger and Schall, 2014) that comprises linear and bi-linear regression models as special cases. The new regression model is bi-phasic, but allows for a smooth transition between the two rates of decline in log(CFU) count. The regression model approximates bi-exponential regression models, but is more flexible in the sense that it allows for terminal rates of decline to be greater than initial rates of decline. The model is implemented as a Bayesian non-linear mixed effects (NLME) regression model, fitted jointly to the data of all patients from a trial. Statistical inference about the mean EBA of TB treatments is based on the Bayesian NLME regression model. The posterior predictive distribution of relevant slope parameters of the Bayesian NLME regression model provides insight into the nature of the EBA of TB treatments; specifically, the posterior predictive distribution allows one to judge whether treatments are associated with mono-linear or bi-linear decline of log(CFU) count, and whether log(CFU) count is predicted initially to decrease fast, followed by a slower rate of decrease, or *vice versa.*

Section 2 provides general aspects which should be considered when fitting regression models to CFU data. In Section 3 below we present and derive the non-linear regression model, and in Section 4, we describe its implementation as a Bayesian NLME regression model. In addition, the extension of the methodology to quantitative liquid

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culture data is discussed in Section 5. Section 6.1 summarises the results of an extensive empirical investigation of the suitability of the model fitted on a by-patient basis, and Section 6.2 is devoted to an application of the methodology to the data of a recently published EBA study.

2 GENERAL CONSIDERATIONS

When fitting regression models to CFU data, the following three important aspects, namely the identification of censored data, handling of sparse data profiles of individual patients, and outliers should be considered:

- **Variable**: Provided that two CFU plate counts are associated with a given sputum sample from two different plates, the log(CFU) counts are calculated as follows:

$$\log(\text{CFU}) \text{ count} = \log_{10}(\text{Mean of two CFU plate counts} \times 20 \times 10^{\text{dilution}})$$

  In the above formula, the factor 20 compensates for the dilution of the specimens during the culture process, converting the result back to the actual CFU count per mL.

- **Censored data**: CFU counts of zero must be identified and confirmed to be "genuine", i.e. genuine zero counts must be distinguished from missing CFU values, or from contaminated or otherwise invalid data. Genuine zero counts are valid data and should preferably be included in the analysis as censored observations (Rustomjee et al., 2008). Here, zero CFU counts will be specified as a left censored value of 1. <u>Rationale:</u> The smallest possible CFU count above zero is 1 for counts from the one plate and zero for counts from the other plate with zero dilution,

leading to a calculated log(CFU) count of:

$$\log(\text{CFU}) \text{ count} = \log_{10}([\{0 + 1\}/2] \times 20 \times 10^0) = \log_{10}(10) = 1$$

log(CFU) counts associated with CFUs which are too numerous to count (TNTC) should preferably be right censored at the corresponding upper limit of quantification (ULOQ).

- **Sparse data**: When the data for a given patient is sparse, several problems might occur when fitting the regression model to the data of individual patients.

  - Over-fit of the regression model: It might be inappropriate to fit two slope parameters when there are only 4 or 5 data points.

  - Slope parameters cannot be identified: When data are available only either in the early part or in late part of the study period, it might not be possible to identify and estimate both slope parameters.

  - The node cannot be identified: If the data in the middle of the study period are missing, the node parameter cannot be identified (which can imply that the slope parameters cannot be identified).

  - Convergence problems when trying to fit the regression model.

- **Outliers**: Outliers in log(CFU) counts might be present in the data due to erroneous sampling or reporting of the data, or such values might be true observations, but of extreme nature. As suggested by Gillespie et al. (2002), when individual fitting of the regression function to log(CFU) count is of concern, it is important to exclude implausible data points, i.e. data points causing irregularities in the pattern of an individual patient's CFU counts over time (which do not adhere to the expected biologic pattern of CFU counts over time). Such outliers may produce

unreliable parameter estimates, which may subsequently jeopardise the validity of the statistical inference of EBA.

It is therefore certain that statistical inferences based on regression modelling of CFU count over time need be robust to the aspects listed above.

3 LINEAR, BI-LINEAR AND BI-PHASIC REGRESSION MODEL

In this section, we propose a bi-phasic non-linear regression model for log(CFU) count versus time data (Burger and Schall, 2014). We start with a regression model with constant rate of change (mono-exponential or log-linear regression model), and then generalise to a bi-linear regression model incorporating two rates of change (initial and late). Accordingly, we derive a bi-phasic regression model allowing for smooth transition from the first to the second phase.

3.1 Constant Rate of Change: Linear Regression Model

In the following, let $y = y(t)$ be the CFU count at time t, and similarly, let $\mu = \mu(t)$ denote the expected CFU count at time t. If we assume that the rate of change in expected CFU count is proportional to μ, we obtain the following well-known differential equation:

$$\frac{d\,\mu}{d\,t} = -\lambda^* \cdot \mu \tag{3}$$

Here $\lambda^* > 0$ is the proportionality constant and characterises the rate of decrease. From Equation (3), it follows that:

$$\frac{1}{\mu}\,d\mu = -\lambda^*\,dt \tag{4}$$

Integrating both sides of Equation (4), we have $\int \frac{1}{\mu} d\mu = -\int \lambda^* dt$ with solution:

$$\ln(\mu) = -\int \lambda^* dt = \alpha^* - \lambda^* \cdot t \tag{5}$$

where $\ln(\cdot)$ is the natural logarithm. Equivalently to Equation (5), we can write:

$$\mu = e^{\alpha^*} \cdot e^{-\lambda^* \cdot t} \tag{6}$$

Based on Equation (6), we can postulate the following multiplicative mono-exponential regression model for y, namely:

$$y = e^{\alpha^*} \cdot e^{-\lambda^* \cdot t} \cdot e^{\varepsilon}$$

where e^{ε} is a multiplicative error term at time t. However, often CFU counts y are transformed logarithmically before model fitting, which leads to the log-linear regression model:

$$\log(y) = \alpha - \lambda \cdot t + \varepsilon \tag{7}$$

where $\log(y) = \log_{10}(y)$ is, by convention for this type of data, the logarithm to the base of 10, and therefore $\alpha = \alpha^* / \ln(10)$ (intercept parameter) and $\lambda = \lambda^* / \ln(10)$ (slope parameter). In our experience, after log-transformation, the variance of $\log(\text{CFU})$ count over time is stable, so that the assumption of constant variance for the residual term ε seems appropriate.

3.2 Variable Rate of Change: Bi-Linear and Bi-Phasic Regression Models

As mentioned above, the majority of $\log(\text{CFU})$ count versus time profiles over 14 days of treatment is bi-phasic. If this is the case, the rate of change in $\log(\text{CFU})$ count itself

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changes over time. In general, if we allow λ^* in Equation (3) to be a function of time, namely $\lambda^*(t)$, then Equation (5) becomes $\ln(\mu) = -\int \lambda^*(t)\,dt$, or equivalently, in terms of the logarithm to the base 10:

$$\log(\mu) = -\int \lambda(t)\,dt \tag{8}$$

where $\lambda(t) = \lambda^*(t)/\ln(10)$.

3.2.1 Step Function: Bi-Linear Regression Model

When $\lambda(t)$ in Equation (8) is a step function (see Figure 3.1a), we have:

$$
\begin{aligned}
\lambda(t) &= \lambda_1; & t \leq \kappa \\
\lambda(t) &= \lambda_2; & t > \kappa
\end{aligned}
\tag{9}
$$

Then:

$$
\begin{aligned}
\log(\mu) &= \alpha - \lambda_1 \cdot t; & t \leq \kappa \\
\log(\mu) &= \alpha + (\lambda_2 - \lambda_1) \cdot \kappa - \lambda_2 \cdot t; & t > \kappa
\end{aligned}
$$

which leads to the conventional bi-linear regression model for log(CFU) count, namely:

$$
\begin{aligned}
\log(y) &= \alpha - \lambda_1 \cdot t + \varepsilon; & t \leq \kappa \\
\log(y) &= \alpha + (\lambda_2 - \lambda_1) \cdot \kappa - \lambda_2 \cdot t + \varepsilon; & t > \kappa
\end{aligned}
\tag{10}
$$

Here, the parameter α and κ is the intercept and node parameter of the regression curve, respectively, and the slopes λ_1 and λ_2 characterise the linear decline on or before the node ($t \leq \kappa$) and after the node ($t > \kappa$), respectively.

Lastly, we note it is convenient to write the regression model in Equation (10) in terms of the parameters $\beta_1 = (\lambda_1 + \lambda_2)/2$ and $\beta_2 = (\lambda_2 - \lambda_1)/2$, which are, respectively, the average of and half the difference between the two rate constants λ_1 and λ_2. Then Equation (10) becomes:

$$
\begin{aligned}
\log(y) &= \alpha - \beta_1 \cdot t + \beta_2 \cdot t + \varepsilon; & t \leq \kappa \\
\log(y) &= \alpha - \beta_1 \cdot t - \beta_2 \cdot (t - 2\kappa) + \varepsilon; & t > \kappa
\end{aligned}
\tag{11}
$$
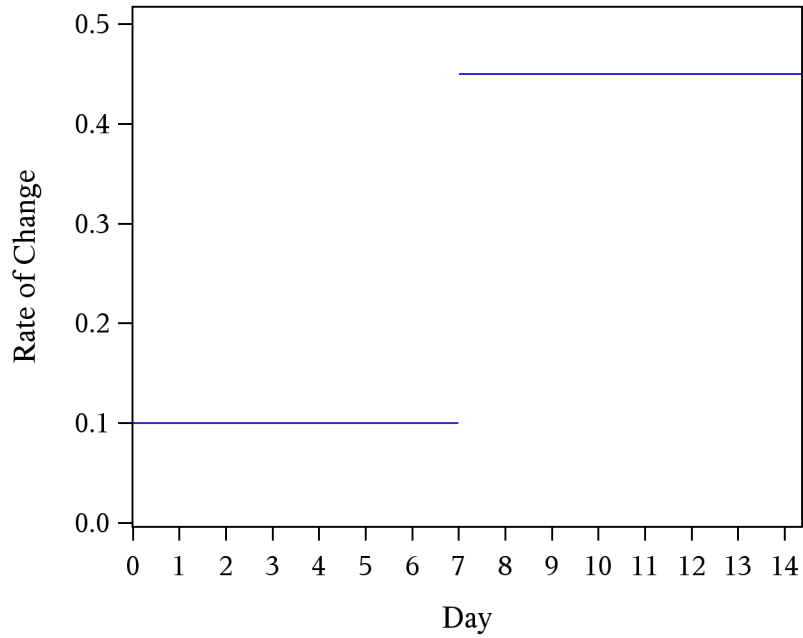
3.2.2 Hyperbolic Tangent Function: Bi-Phasic Regression Model

As has been pointed out by Jindani et al. (2003), the switch from one rate of decline in log(CFU) count to another might be smooth, rather than abrupt as is implied with by the bi-linear regression model in Equation (10). In order to model a smooth transition, we can use a monotonic function that interpolates between the early rate of decline, λ_1, and the late rate of decline, λ_2. For example, a class of such functions is formed by linear transformations of cumulative distribution functions (Seber and Wild, 1989).

In the following, we model $\lambda(t)$ using the hyperbolic tangent function:

$$
\lambda(t) = \frac{\lambda_1 + \lambda_2}{2} + \frac{\lambda_2 - \lambda_1}{2} \cdot \frac{e^{\frac{t-\kappa}{\gamma}} - e^{-\frac{t-\kappa}{\gamma}}}{e^{\frac{t-\kappa}{\gamma}} + e^{-\frac{t-\kappa}{\gamma}}}
\tag{12}
$$

The hyperbolic tangent function in Equation (12), shown in Figure 3.1b, is essentially a smooth version of the step function in Equation (9). For small t, $\lambda(t)$ tends to λ_1, i.e. $\lim_{t \to 0} \lambda(t) = \lambda_1$, and similarly, for large t, the function $\lambda(t)$ tends to λ_2, i.e. $\lim_{t \to \infty} \lambda(t) = \lambda_2$. Furthermore, $\lambda(\kappa) = (\lambda_1 + \lambda_2)/2$, so that κ can be viewed as the "node" of the function $\lambda(t)$. Lastly, the parameter γ governs the "smoothness" of the transition from rate λ_1 to rate λ_2. With $\lambda(t)$ as in Equation (12), we obtain $\mu = \mu(t)$ as

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Figure 3.1: Example Plot of Rate of Change in Expected log(CFU) Count ($\lambda(t)$) Over Time (Days)

(a) Step Function
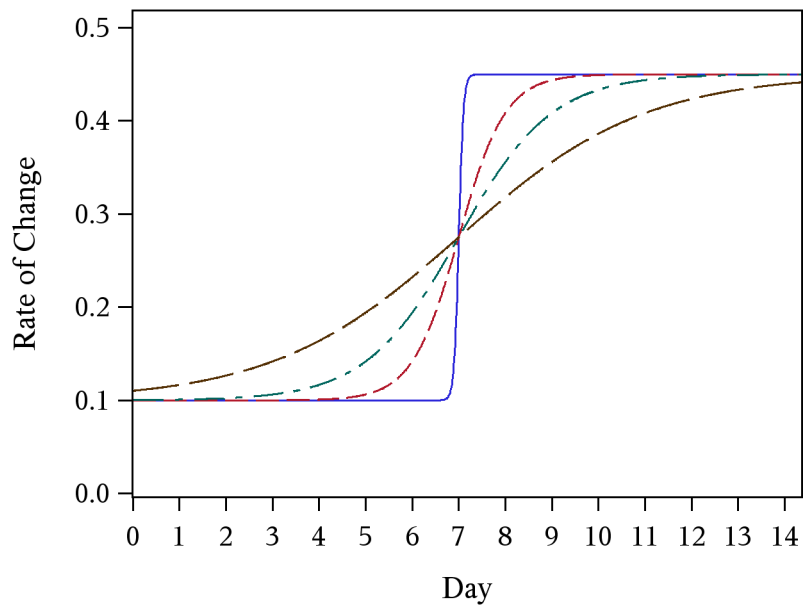
(b) Hyperbolic Tangent Function

the integral in Equation (8), namely:

$$\log(\mu) = \alpha - \frac{\lambda_1 + \lambda_2}{2} \cdot t - \frac{\lambda_2 - \lambda_1}{2} \cdot \gamma \cdot \ln \left(\frac{e^{\frac{t-\kappa}{\gamma}} + e^{-\frac{t-\kappa}{\gamma}}}{e^{\frac{\kappa}{\gamma}} + e^{-\frac{\kappa}{\gamma}}} \right)$$

Thus we have the following bi-phasic non-linear regression model for $\log(y)$:

$$\log(y) = \alpha - \frac{\lambda_1 + \lambda_2}{2} \cdot t - \frac{\lambda_2 - \lambda_1}{2} \cdot \gamma \cdot \ln \left(\frac{e^{\frac{t-\kappa}{\gamma}} + e^{-\frac{t-\kappa}{\gamma}}}{e^{\frac{\kappa}{\gamma}} + e^{-\frac{\kappa}{\gamma}}} \right) + \varepsilon \tag{13}$$

Note that, for small t (and small γ relative to κ), the term $e^{\frac{t-\kappa}{\gamma}}$ tends to zero, while the term $e^{-\frac{t-\kappa}{\gamma}}$ becomes large. Thus, for small t ($t \leq \kappa$), $\log(\mu)$ declines approximately linearly with slope $-\lambda_1$. *Vice versa*, for large t, the term $e^{\frac{t-\kappa}{\gamma}}$ becomes large, while the term $e^{-\frac{t-\kappa}{\gamma}}$ tends to 0. Thus, for large t, ($t \geq \kappa$), $\log(\mu)$ declines approximately linearly with slope $-\lambda_2$.

In summary, the regression model in Equation (13) is a "smooth" version of the bi-linear regression model in Equation (10). (In fact, the regression model in Equation (10) is a special case of the regression model in Equation (13) when $\gamma \to 0$.) The parameters λ_1 and λ_2 can therefore be interpreted as the "early" and "late" rates of decline, respectively, while the parameter α is the intercept of the regression curve. Furthermore, γ characterises the "smoothness" of the transition from the early to the terminal decay curve, and κ is the node parameter. Furthermore, for small t (when $\lambda_1 > \lambda_2 > 0$), the variable y (i.e. CFU count on the original scale) is approximated by an exponential function $C_1 \cdot e^{-\lambda_1 \cdot t}$, where $C_1 = \exp\left(\alpha - \frac{\lambda_2 - \lambda_1}{2} \cdot \left[\kappa - \gamma \cdot \ln\left\{e^{\frac{\kappa}{\gamma}} + e^{-\frac{\kappa}{\gamma}}\right\}\right]\right)$ and for large t, the variable y is approximated by an exponential function $C_2 \cdot e^{-\lambda_2 \cdot t}$, where $C_2 = \exp\left(\alpha + \frac{\lambda_2 - \lambda_1}{2} \cdot \left[\kappa + \gamma \cdot \ln\left\{e^{\frac{\kappa}{\gamma}} + e^{-\frac{\kappa}{\gamma}}\right\}\right]\right)$. In that sense, the regression model in Equation (13) approximates bi-exponential regression models.

Lastly, when $\beta_1 = (\lambda_1 + \lambda_2)/2$ and $\beta_2 = (\lambda_2 - \lambda_1)/2$, Equation (13) becomes:

$$\log(y) = \alpha - \beta_1 \cdot t - \beta_2 \cdot \gamma \cdot \ln\left(\frac{e^{\frac{t-\kappa}{\gamma}} + e^{-\frac{t-\kappa}{\gamma}}}{e^{\frac{\kappa}{\gamma}} + e^{-\frac{\kappa}{\gamma}}} \right) + \varepsilon \tag{14}$$

The regression models in Equation (13) and Equation (14) can be fitted to the log(CFU) count versus time data of individual patients using maximum likelihood (ML) estimation (similar to conventional "by-patient" regression modelling by Diacon et al. (2012a, 2013)). Relevant EBA parameters can accordingly be estimated for each patient based on these model fits.

It should be noted that the regression models in Equation (13) and Equation (14) are similar to models proposed by Bacon and Watts (1971); Griffiths and Miller (1973); Ratkowsky (1983); Grossman et al. (1999), which also have two intersecting line segments as a limiting case; these models comprise parameterisations different to our proposed model.

4   BAYESIAN FIT OF REGRESSION MODELS

4.1   Model 1: <u>Bi-Phasic – Student t Errors and 'Default' Wishart Priors</u>

We propose a bi-phasic hierarchical Bayesian NLME regression model for log(CFU) count versus time (Burger and Schall, 2014), fitted jointly to the data of all patients from a given trial.

We start by specifying an NLME regression model for the log(CFU) counts. Let y_{ijk} be the CFU count for patient $i = 1, \ldots, N_j$ in treatment group $j = 1, \ldots, J$ at time-point $k = 1, \ldots, K_{ij}$, and let t_{ijk} be the corresponding measurement time. Then,

based on Equation (14), we write the following NLME regression model:

$$\log(y_{ijk}) = \alpha_{ij} - \beta_{1ij} \cdot t_{ijk} - \beta_{2ij} \cdot \gamma_{ij} \cdot \ln\left(\frac{e^{\frac{t_{ijk}-\kappa_{ij}}{\gamma_{ij}}} + e^{-\frac{t_{ijk}-\kappa_{ij}}{\gamma_{ij}}}}{e^{\frac{\kappa_{ij}}{\gamma_{ij}}} + e^{-\frac{\kappa_{ij}}{\gamma_{ij}}}}\right) + \varepsilon_{ijk} \qquad (15)$$

The parameters of the regression model in Equation (15) are analogous to those of the "by-patient" regression model in Equation (14).

The subsections below provide a full specification of the random effects and prior distributions of the regression model in Equation (15).

Random Effects

The vectors $\boldsymbol{\mu_{ij}} = (\alpha_{ij}, \beta_{1ij}, \beta_{2ij})'$ of intercept and slope parameters are assumed independent across patients (i.e. independent across indices i and j), with tri-variate normal distributions as follows:

$$\boldsymbol{\mu_{ij}} \sim N(\boldsymbol{\mu_j}, \boldsymbol{\Omega_{\mu j}}) \qquad (16)$$

In Equation (16), $\boldsymbol{\mu_j} = (\alpha_j, \beta_{1j}, \beta_{2j})'$ are vectors of mean intercepts and slopes, and $\boldsymbol{\Omega_{\mu j}}$ are the associated covariance matrices, namely:

$$\boldsymbol{\Omega_{\mu j}} = \begin{bmatrix} \sigma^2_{\alpha j} & \text{Cov}_j(\alpha_{ij}, \beta_{1ij}) & \text{Cov}_j(\alpha_{ij}, \beta_{2ij}) \\ \text{Cov}_j(\alpha_{ij}, \beta_{1ij}) & \sigma^2_{\beta_1 j} & \text{Cov}_j(\beta_{1ij}, \beta_{2ij}) \\ \text{Cov}_j(\alpha_{ij}, \beta_{2ij}) & \text{Cov}_j(\beta_{1ij}, \beta_{2ij}) & \sigma^2_{\beta_2 j} \end{bmatrix}$$

Furthermore, the parameters κ_{ij} and γ_{ij} are assumed to follow truncated normal distributions, independent of each other, and independent of $\boldsymbol{\mu_{ij}}$, as follows:

$$\begin{aligned} \kappa_{ij} &\sim TN(\kappa_j, \sigma^2_{\kappa j}) \cdot I(L_\kappa \leq \kappa_{ij} \leq U_\kappa) \\ \gamma_{ij} &\sim TN(\gamma_j, \sigma^2_{\gamma j}) \cdot I(L_\gamma \leq \gamma_{ij} \leq U_\gamma) \end{aligned} \qquad (17)$$

18

In Equation (17), $I(x)$ denotes an indicator function taking the value 1 if x is true, and 0 otherwise, and L_κ, U_κ, L_γ and U_γ are the pre-specified lower bound and upper bound for parameters κ_{ij} and γ_{ij}, respectively.

Finally, the residuals ε_{ijk} are assumed to follow independent Student t distributions, independent of $\boldsymbol{\mu_{ij}}$, κ_{ij} and γ_{ij}, as follows:

$$\varepsilon_{ijk} \sim T(0, \sigma_{\varepsilon j}^2, v_j) \tag{18}$$

where $\sigma_{\varepsilon j}^2$ and v_j are scale parameters and degrees of freedom, respectively, from the corresponding Student t distribution. The specification of the Student t distribution can accommodate heavily tailed residual errors which, in this regard, is more flexible than the normal distribution.

A subset of CFU counts might be reported as zero or "no count" values. Genuine zero counts will typically occur when, for a given patient profile, CFU counts are observed over time to decline to near zero values, just prior to observing one or more zero counts. Thus, genuine zero counts will typically occur towards the end of a CFU count versus time profile. When regressing log(CFU) count against time using Equation (15), the log(CFU) counts corresponding to zero count can be specified as a left-censored value of 1 (formally, $\log(y_{ijk}) < 1$) (Rustomjee et al., 2008).

Prior Distributions

In order to complete the Bayesian specification of the NLME regression model described above, proper but vague prior distributions are assigned to all unknown parameters of the NLME regression model.

Firstly, multivariate normal and Wishart prior distributions are specified, respec-

tively, for $\boldsymbol{\mu_j}$ and $\boldsymbol{\Omega_{\mu j}^{-1}}$ in Equation (16), namely:

$$\boldsymbol{\mu_j} \sim N(\mathbf{0}, 10^4 \times \boldsymbol{I_3}) \tag{19}$$

$$\boldsymbol{\Omega_{\mu j}^{-1}} \sim W(3, 3 \times \boldsymbol{R_j}) \tag{20}$$

where $\mathbf{0} = (0,0,0)'$ and $\boldsymbol{I_3}$ denotes the 3×3 identity matrix. $\boldsymbol{R_j}$ represent 3×3 inverse scale matrices.

One challenge is the choice of an appropriate prior distribution for the covariance matrix of the vectors of intercept and slope parameters $\boldsymbol{\mu_{ij}}$, i.e. $\boldsymbol{\Omega_{\mu j}}$. We used the methodology by Kass and Natarajan (2006), referred to as the "default" Wishart prior, for choosing $\boldsymbol{R_j}$. This methodology relates to the choice of $\boldsymbol{R_j}$ in the application of generalised linear mixed effects regression modelling, and is derived from the data directly (hence, the resulting posterior distribution does make double use of the data). The inverse scale matrix $\boldsymbol{R_j}$ is derived by selecting the weight which the mean of the "shrinkage" prior, i.e. $\mathbf{0}$, should contribute towards its posterior (where "shrinkage" represents $\boldsymbol{\mu_{ij}} - \boldsymbol{\mu_j}$). Under the assumption that the node and smoothness parameters are fixed at $\kappa_p = (U_\kappa + L_\kappa)/2$ and $\gamma_p = (U_\gamma + L_\gamma)/2$, respectively (which are the prior mean for κ_j and γ_j, respectively (see below)), the regression model with normally distributed errors in Equation (15) reduces to a linear mixed effects regression model, for which $\boldsymbol{R_j}$ are derived as follows:

$$\boldsymbol{R_j} = c \cdot \left(\frac{1}{N_j \cdot \tilde{\sigma}_{\varepsilon j}^2} \sum_{i=1}^{N_j} \boldsymbol{Z_{ij}'} \cdot \boldsymbol{Z_{ij}} \right)^{-1} \tag{21}$$

where $\tilde{\sigma}_{\varepsilon j}^2$ are the ML estimates of $\sigma_{\varepsilon j}^2$ when assuming the regression model is homogeneous across all patients (i.e. disregarding random effects such that $\alpha_{ij} = \alpha_j$, $\beta_{1ij} = \beta_{1j}$

and $\beta_{2ij} = \beta_{2j}$). The matrices $\boldsymbol{Z_{ij}}$ are defined as follows:

$$
\boldsymbol{Z_{ij}} =
\begin{bmatrix}
1 & -t_{ij1} & -\gamma_p \cdot \ln\left(\dfrac{e^{\frac{t_{ij1}-\kappa_p}{\gamma_p}}+e^{-\frac{t_{ij1}-\kappa_p}{\gamma_p}}}{e^{\frac{\kappa_p}{\gamma_p}}+e^{-\frac{\kappa_p}{\gamma_p}}} \right) \\
\vdots & \vdots & \vdots \\
1 & -t_{ijk} & -\gamma_p \cdot \ln\left(\dfrac{e^{\frac{t_{ijk}-\kappa_p}{\gamma_p}}+e^{-\frac{t_{ijk}-\kappa_p}{\gamma_p}}}{e^{\frac{\kappa_p}{\gamma_p}}+e^{-\frac{\kappa_p}{\gamma_p}}} \right) \\
\vdots & \vdots & \vdots \\
1 & -t_{ijK_{ij}} & -\gamma_p \cdot \ln\left(\dfrac{e^{\frac{t_{ijK_{ij}}-\kappa_p}{\gamma_p}}+e^{-\frac{t_{ijK_{ij}}-\kappa_p}{\gamma_p}}}{e^{\frac{\kappa_p}{\gamma_p}}+e^{-\frac{\kappa_p}{\gamma_p}}} \right)
\end{bmatrix}
$$

We used $c = 2.5$, causing the mean of the 'shrinkage" prior, i.e. $\boldsymbol{0}$, to have little contribution towards its posterior. The choice of $c = 2.5$ is equivalent to setting the interval between the lowest and highest possible values for the relative contribution matrix of the mean of the "shrinkage" prior (to its posterior) to 28.6%.

The parameters κ_j, γ_j, $\sigma^2_{\kappa j}$ and $\sigma^2_{\gamma j}$ (see Equation (17)) are assumed to follow uniform prior distributions, namely $\kappa_j \sim U(L_\kappa, U_\kappa)$, $\gamma_j \sim U(L_\gamma, U_\gamma)$, $\sigma^2_{\kappa j} \sim U(L_{\sigma^2_{\kappa_j}}, U_{\sigma^2_{\kappa_j}})$ and $\sigma^2_{\gamma j} \sim U(L_{\sigma^2_{\gamma_j}}, U_{\sigma^2_{\gamma_j}})$, where $L_{\sigma^2_\kappa}, U_{\sigma^2_\kappa}, L_{\sigma^2_\gamma}, U_{\sigma^2_\gamma}$ are the pre-specified lower bound and upper bound for parameters $\sigma^2_{\kappa j}$ and $\sigma^2_{\gamma j}$, respectively.

Finally, the scale parameters $\sigma^2_{\varepsilon j}$ and degrees of freedom v_j in Equation (18) are respectively assigned inverse gamma prior distributions, namely $\sigma^2_{\varepsilon j} \sim IG(10^{-4}, 10^{-4})$, and uniform prior distributions, namely $v_j \sim U(2, 100)$.

For a typical 14-day EBA study, the hyper parameters of the prior distributions can be chosen as follows: $L_\kappa = 2$, $U_\kappa = 11$ (to avoid over-fit of the first few and last few observations over time), $L_\gamma = 0.1$, $U_\gamma = 2$ (allowing for smooth transition between a few successive data points), $L_{\sigma^2_\kappa} = 0.01$, $U_{\sigma^2_\kappa} = 30$, $L_{\sigma^2_\gamma} = 0.01$ and $U_{\sigma^2_\gamma} = 5$ (providing weakly informative prior distributions for the scale parameters $\sigma^2_{\kappa j}$ and $\sigma^2_{\gamma j}$).

4.2 Model 2: Bi-Phasic – Student t Errors and 'Frequentist' Wishart Priors

To assess the sensitivity of results to the choice of $\boldsymbol{R_j}$, we fitted Model 1 as a linear mixed effects regression model under the assumption that the node and smoothness parameters (i.e. κ_{ij}, κ_j, γ_{ij} and γ_j) are fixed at $(U_\kappa + L_\kappa)/2$ and $(U_\gamma + L_\gamma)/2$, respectively. We calculated the "frequentist" estimates for $\boldsymbol{\Omega_{\mu j}}$ via ML estimation (using the SAS® procedure PROC NLMIXED) to serve as $\boldsymbol{R_j}$.

4.3 Model 3: Bi-Phasic – Normal Errors and 'Default' Wishart Priors

Model 1 can incorporate the assumption that the residual errors follow normal distributions (i.e. instead of Student t distributed residual errors), i.e. $\varepsilon_{ijk} \sim N(0, \sigma^2_{\varepsilon j})$, where $\sigma^2_{\varepsilon j}$ are the corresponding residual variances following inverse gamma prior distributions, namely $\sigma^2_{\varepsilon j} \sim IG(10^{-4}, 10^{-4})$.

4.4 Model 4: Bi-Phasic – Normal Errors and 'Frequentist' Wishart Priors

The sensitivity of results to the choice of $\boldsymbol{R_j}$ in Model 3 can be assessed using the "frequentist" approach specified for Model 2.

4.5 Model 5: Bi-Linear – Student t Errors and 'Default' Wishart Priors

Based on Equation (11), we can postulate the following bi-linear mixed effects regression model:

$$\log(y_{ijk}) = \alpha_{ij} - \beta_{1ij} \cdot t_{ijk} + (-1)^{J_{ijk}+1} \cdot \beta_{2ij} \cdot t_{ijk} + 2\,(J_{ijk} - 1) \cdot \beta_{2ij} \cdot \kappa_{ij} + \varepsilon_{ijk} \quad (22)$$

where $J_{ijk} = 1 + \text{step}\,(t_{ijk} - \kappa_{ij})$, and $\text{step}\,(x)$ denotes a function taking the value 0 if $x \leq 0$, and 1 otherwise. The parameters of the regression model in Equation (22) are analogous to those of the "by-patient" regression model in Equation (11), and the

specification of its random effects and prior distributions are similar to those of Model 1.

4.6 Model 6: <u>Bi-Linear – Normal Errors and 'Default' Wishart Priors</u>

Model 5 can incorporate the assumption that the residual errors follow normal distributions (i.e. instead of Student t distributed residual errors).

4.7 Model 7: <u>Mono-Linear – Student t Errors and 'Default' Wishart Priors</u>

The conventional linear mixed effects regression model can we written as follows:

$$\log(y_{ijk}) = \alpha_{ij} - \lambda_{ij} \cdot t_{ijk} + \varepsilon_{ijk} \tag{23}$$

The parameters of the regression model in Equation (23) are analogous to those of the "by-patient" regression model in Equation (7), and the specification of its random effects and prior distributions are similar to those of Model 1.

4.8 Model 8: <u>Mono-Linear – Normal Errors and 'Default' Wishart Priors</u>

Model 7 can incorporate the assumption that the residual errors follow normal distributions (i.e. instead of Student t distributed residual errors).

4.9 Posterior Predictive Distributions

The posterior predictive distribution of relevant slope parameters of the Bayesian NLME regression model provides insight into the nature of the EBA of TB treatments; specifically, the posterior predictive distributions of β_{2j} allow one to judge whether treatments are associated with mono-linear or bi-phasic decline of log(CFU) count (depending on whether a future β_{2j} is likely to be close to or substantially different from zero), and whether log(CFU) count initially decreases fast, followed by a slower rate of decrease

(if a future β_{2j} is likely to be negative), or *vice versa* (if a future β_{2j} is likely to be positive). The simulation of the posterior predictive distribution of the future regression slopes β_{2fj} (where the subscript f stands for "future patient") can be implemented in a straightforward manner using the Markov Chain Monte Carlo (MCMC) output of the Gibbs sampling algorithm of the joint posterior distribution of the regression model parameters.

4.10   Model Selection and Model Checking

Alternative NLME regression models can be explored via various Bayesian model selection tools, and may be fitted to assess:

- Alternative shapes of the log(CFU) count versus time profiles, e.g. assuming a linear, bi-linear or bi-phasic relationship between log(CFU) count and time.

- The sensitivity of results to the choice of prior distributions.

- Alternative distributions for random effects and residuals (error terms).

In order to check our primary model (Model 1; Section 4.1), and to assess the aspects listed above, we fitted the seven additional models (with alternative Bayesian specifications) specified in Section 4.2 through Section 4.8. The fit of each of the models was checked using conditional posterior ordinates (CPOs) and their reciprocals (ICPOs). Some detail is included in the appendix.

Two methods for discriminating between various regression models were considered: The deviance information criterion (DIC) (Spiegelhalter et al., 2002) and Bayes factors (Kass and Raftery, 1995).

Deviance Information Criterion

The DIC is a model adequacy and goodness of fit measure, and is defined for Model M as follows:

$$\text{DIC}(M) = 2\overline{\text{D}(\boldsymbol{\theta_m}, M)} - \text{D}(\bar{\boldsymbol{\theta}}_{\boldsymbol{m}}, M) = \text{D}(\bar{\boldsymbol{\theta}}_{\boldsymbol{m}}, M) + 2p_m \tag{24}$$

where $\boldsymbol{\theta_m}$ is a $d_m \times 1$ vector of model parameters, $\boldsymbol{y}$ is an $n \times 1$ vector of observed data, $\text{D}(\boldsymbol{\theta_m}, M) = -2\ln(f(\boldsymbol{y}|\boldsymbol{\theta_m}, M))$ is the conventional deviance measure (i.e. minus twice the log-likelihood), $\bar{\boldsymbol{\theta}}_{\boldsymbol{m}}$ and $\overline{\text{D}(\boldsymbol{\theta_m}, M)}$ are the mean of the posterior distribution of $\boldsymbol{\theta_m}$ and $\text{D}(\boldsymbol{\theta_m}, M)$, respectively, and $p_m = \overline{\text{D}(\boldsymbol{\theta_m}, M)} - \text{D}(\bar{\boldsymbol{\theta}}_{\boldsymbol{m}}, M)$ is the number of "effective" parameters.

The quantity $\text{DIC}(M)$ is therefore a measure which takes both goodness of fit and complexity of Model M into account, and is more appropriate to assess the predictability of random effects in Model M (Spiegelhalter et al., 2003). The model with the smallest DIC is considered to fit the data more appropriately. However, the DIC measure may be unreliable in cases where $\bar{\boldsymbol{\theta}}_{\boldsymbol{m}}$ is an unreliable estimator of $\boldsymbol{\theta_m}$ (Ntzoufras, 2009).

Bayes Factors

When comparing Model M_0 and Model M_1, based on the posterior probability of each of the models given the data, the Bayes factor in favour of M_0 is defined as follows:

$$B_{01} = \frac{f(\boldsymbol{y}|M_0)}{f(\boldsymbol{y}|M_1)} \tag{25}$$

where $\boldsymbol{y}$ is an $n \times 1$ vector of observed data, and $f(\boldsymbol{y}|M_0)$ and $f(\boldsymbol{y}|M_1)$ are the marginal likelihoods of $\boldsymbol{y}$ under Model M_0 and Model M_1, respectively.

Unlike the DIC, Bayes factors do not explicitly include a term that penalises model complexity, but rather incorporates the latter in the marginal likelihood of a given model

automatically (Ward, 2008). Furthermore, the DIC compares models conditional on their model parameters, whereas the Bayes factors compare models on a marginal basis.

In the case of NLME regression modelling, the marginal likelihoods in Equation (25) need to be approximated. The Laplace-Metropolis approximation, in its general form, for $\ln(f(\boldsymbol{y}|M))$ is given by the following expression (Ntzoufras, 2009):

$$\ln(\hat{f}(\boldsymbol{y}|M)) = \frac{1}{2}d_m \ln(2\pi) + \frac{1}{2}\ln|R_{\boldsymbol{\theta_m}}| + \sum_{j=1}^{d_m}\ln(s_j) + \sum_{i=1}^{n}\ln(f(y_i|\bar{\boldsymbol{\theta}}_{\boldsymbol{m}}, M)) + \sum_{j=1}^{d_m}\ln(f(\bar{\theta}_{mj}|M))$$

(26)

where $\bar{\theta}_{mj}$ and s_j are the mean and standard deviation (SD), respectively, of the posterior distribution of θ_{mj}, and $|R_{\boldsymbol{\theta_m}}|$ is the determinant of the $d_m \times d_m$ correlation matrix of the posterior distribution of $\boldsymbol{\theta_m}$. In mixed effects models, the calculation of the Laplace-Metropolis marginal likelihood requires that the random effects included in each patient's likelihood function should be integrated out (Lewis and Raftery, 1997). The 5 random effects (see Model 1) for each patient were marginalised using the multidimensional integration library R2Cuba of the R project (R Core Team, 2014; Hahn et al., 2013). The Laplace-Metropolis approximation in Equation (26) is based on asymptotic theory of the normal distribution, and works well for symmetric posterior distributions of $\boldsymbol{\theta_m}$ (Ntzoufras, 2009).

4.11 Computational Issues

The OpenBUGS software (Version 3.2.2) is used to implement the MCMC Gibbs sampling algorithm to draw samples from the joint posterior distribution of the model parameters (Gelfand and Smith, 1990; Gilks et al., 1996; Lunn et al., 2009). OpenBUGS can be downloaded for free from URL `http://www.openbugs.net/w/Downloads`.

Due to the high dimensional nature of NLME regression models, by-patient parameter estimates, obtained from regression fits (such as Equation 14) for each patient

individually (using SAS® procedure PROC NLMIXED), were used as starting values for the random effects. The posterior samples were thinned to reduce the autocorrelation among posterior samples. Graphical convergence diagnostics, such as iteration and autocorrelation plots, and the Brooks-Gelman-Rubin statistic (Brooks and Gelman, 1998) for two parallel chains, were used to monitor convergence of posterior samples. Dispersed starting values for the second chain were provided to ensure convergence of the two respective chains. Multidimensional integrals (for calculation of Laplace-Metropolis marginal likelihoods) were calculated using libraries available in the R project.

5   EXTENSION TO QUANTITATIVE LIQUID CULTURE

Liquid culture is generally considered more sensitive than solid culture. In liquid culture, the opportunity to count colonies of bacteria is not available, but the time it takes for growth in liquid culture to register as a positive readout (time to positivity, or TTP) is inversely related to the bacterial load of such cultures (Diacon et al., 2012b). Liquid culture can therefore also be reported out quantitatively. Traditionally, serial TTP data have been presented on a linear scale because the measure already incorporates a growth function. A preliminary investigation of TTP data suggested that both TTP and log(TTP) data increase linearly or bi-linearly over time (Diacon et al., 2012a). However, the log-transformed TTP data versus time profiles suggest that the residual variance is constant over the range of fitted values (as opposed to TTP data on the original scale). That is, the logarithm is effective as variance stabilising transformation. We therefore recommend the analysis of TTP data on the logarithmic scale instead.

When fitting regression models to TTP data, the following important aspects, in addition to those applicable to CFU data, should be considered:

- **Variable**: Provided that two TTP values are associated with a given sputum

sample from two different sets, log(TTP) is calculated as follows:

$$\log(\text{TTP}) = \log_{10}(\text{Mean of two TTP values})$$

- **Censored data**: TTP values might be reported as "negative" (i.e., no mycobacterial growth). The manufacturer's recommended incubation time before reporting a result as "negative" is 42 days (equivalently, 1008 hours). Thus the largest possible numeric TTP value that can be observed is 1008 hours, for an incubation time of 1008 hours. When regressing log(TTP) against time, the log(TTP) values reported as "negative" are specified as right censored values. In an ongoing Phase 3 study (REMoxTB), where sputa from approximately 2000 patients were collected serially over 18 months of treatment and follow-up, only 6.8% of the reported positive liquid cultures had TTP values exceeding 600 hours. The censoring time could be chosen to be equal to the incubation time (1008 hours); however, because experience suggests that TTP values reported above 600 hours are rare, the following censoring rule is used: TTP values reported as "negative" should be right-censored at 600 hours, or the maximum TTP value observed in the study, whichever is greater.

The joint Bayesian NLME regression models discussed in Section 4 can be fitted to the log(TTP) versus time data, however, incorporating a slight modification in the sign of the slope parameters, i.e. "$+ \beta_{1ij}$" and "$+ \beta_{2ij}$" instead of "$- \beta_{1ij}$" and "$- \beta_{2ij}$".

6 EMPIRICAL STUDY AND EXAMPLE OF APPLICATION

6.1 Empirical Study

While theoretical considerations may assist in the derivation of a suitable regression model for a certain type of data, the most important requirement for a good regression model is that it should fit the data well. Thus, in deriving a regression model for CFU count, we have started with an empirical study of a large number of log(CFU) count versus time profiles from four EBA trials. The typical shapes of such profiles, identified in the empirical study, confirm observations made previously by other authors and motivate the theoretical derivation of the bi-phasic non-linear regression model proposed in Section 3.2.2.

For the purpose of this empirical study, we have had access to the data from four EBA trials comprising of CFU count versus time profiles of a total of 291 patients. In all four trials, CFU data were collected over a period of 14 days of treatment. Relevant clinical trial characteristics of clinical trial protocol CL001, CL007, CL010 and NC001 are summarised in Table 6.1, including the total number of randomised patients, and the number of randomised patients with complete profiles (data up to Day 14).

The log(CFU) count versus time profiles of all patients with complete profiles were fitted, separately by patient, using the SAS® procedure NLMIXED. Note that we used only patients with complete data profiles since the primary purpose of the empirical study was to judge the adequacy of the proposed bi-phasic model specifically when fitted to 14-day CFU count versus time profiles; naturally, when data profiles are (substantially) shorter than 14 days (e.g. due to a patient dropping out of a trial early) a simple mono-linear model will often be adequate.

Plots of the data together with by-patient fits of the bi-phasic regression model are

included as Figure A.1 through Figure A.21 in the supplementary material. The residuals were assumed to follow independent and identically distributed normal distributions, and the lower and upper bounds of κ and γ were respectively set to $L_\kappa = 2$, $U_\kappa = 11$, $L_\gamma = 0.1$ and $U_\gamma = 2$.

Table 6.1: Characteristics of Clinical Trials Included in Empirical Study

Clinical Trial	Scheduled Sample Days	Treatment Group	N	n
CL001	Daily from Day -2 to Day 8;	TMC207 100 mg	15	12
	Day 10, Day 12, Day 14	TMC207 200 mg	15	13
		TMC207 200 mg	15	13
		TMC207 400 mg	15	14
		Rifafour	8	6
		Total	68	58
CL007	Daily from Day -2 to Day 4;	PA-824 200 mg	15	12
	Day 6, Day 8, Day 10, Day 12,	PA-824 600 mg	15	12
	Day 14	PA-824 1000 mg	16	15
		PA-824 1200 mg	15	11
		Rifafour	8	7
		Total	69	57
CL010	Daily from Day -2 to Day 4;	PA-824 50 mg	15	12
	Day 6, Day 8, Day 10, Day 12,	PA-824 100 mg	15	15
	Day 14	PA-824 150 mg	15	14
		PA-824 200 mg	16	14
		Rifafour	8	8
		Total	69	63

Note: Treatment Group: J = TMC207, J-Z = TMC207 + Pyrazinamide, J-Pa = TMC207 + PA-824, Pa-Z = PA-824 + Pyrazinamide, Pa-Z-M = PA-824 + Pyrazinamide + Moxifloxacin, Rifafour = Rifafour e-275®. N = Total number of randomised patients. n = Number of randomised patients with complete profiles.

Table 6.1: Characteristics of Clinical Trials Included in Empirical Study

Clinical Trial	Scheduled Sample Days	Treatment Group	N	n
NC001	Daily from Day -2 to Day 14	J	15	14
		J -Z	15	12
		J-Pa	15	12
		Pa-Z	15	13
		Pa-Z-M	15	10
		Rifafour	10	8
		Total	85	69
Total		Total	291	247

Note: Treatment Group: J = TMC207, J-Z = TMC207 + Pyrazinamide, J-Pa = TMC207 + PA-824, Pa-Z = PA-824 + Pyrazinamide, Pa-Z-M = PA-824 + Pyrazinamide + Moxifloxacin, Rifafour = Rifafour e-275®. N = Total number of randomised patients. n = Number of randomised patients with complete profiles.

Studying the data profiles, we noted the following (see Table 6.2):

1. Over the profile period of 14 days, the log(CFU) count versus time profiles seem either linear (for the minority of patients: 40 out of 247), or bi-phasic (for the majority of patients: 207 out of 247). For an example of a (near) linear profile, see Figure 6.1a; examples of clearly bi-phasic profiles are given in Figures 6.1b through Figure 6.1d.

2. The rate of decline in log(CFU) count during the initial phase is greater than during the terminal phase for the majority of bi-phasic profiles (e.g. Figure 6.1b); the rate of decline in log(CFU) count during the initial phase is smaller than during the terminal phase for the minority of bi-phasic profiles (e.g. Figure 6.1c).

3. The transition from the first to the second phase is smooth for a minority of bi-phasic profiles (e.g. Figure 6.1d); a bi-linear regression model seems adequate for the majority of bi-phasic profiles (e.g. Figure 6.1b and Figure 6.1c).

4. The average rate of decline in log(CFU) count during the initial phase is for some treatment regimens greater than during the terminal phase. However, for one of the newer compounds under investigation, bedaquiline (TMC207), and for some treatment regimens containing TMC207 in combination with other drugs, the average rate of decline in log(CFU) count during the initial phase is smaller than during the terminal phase.

5. Whatever the respective *average* rates of decline in log(CFU) count for a given treatment regimen, rates of decline both during the initial and late phase exhibit appreciable inter-individual variability; for individual patients, the rate of decline in log(CFU) count during the initial phase might be smaller than during the late phase, even though the respective average rates for the treatment regimen in question might exhibit the reverse relationship.

6. The time-point (node) at which the initial rate of decline changes to the terminal rate of decline exhibits appreciable individual variability (possibly as a result of little information for the estimation of the node parameter).

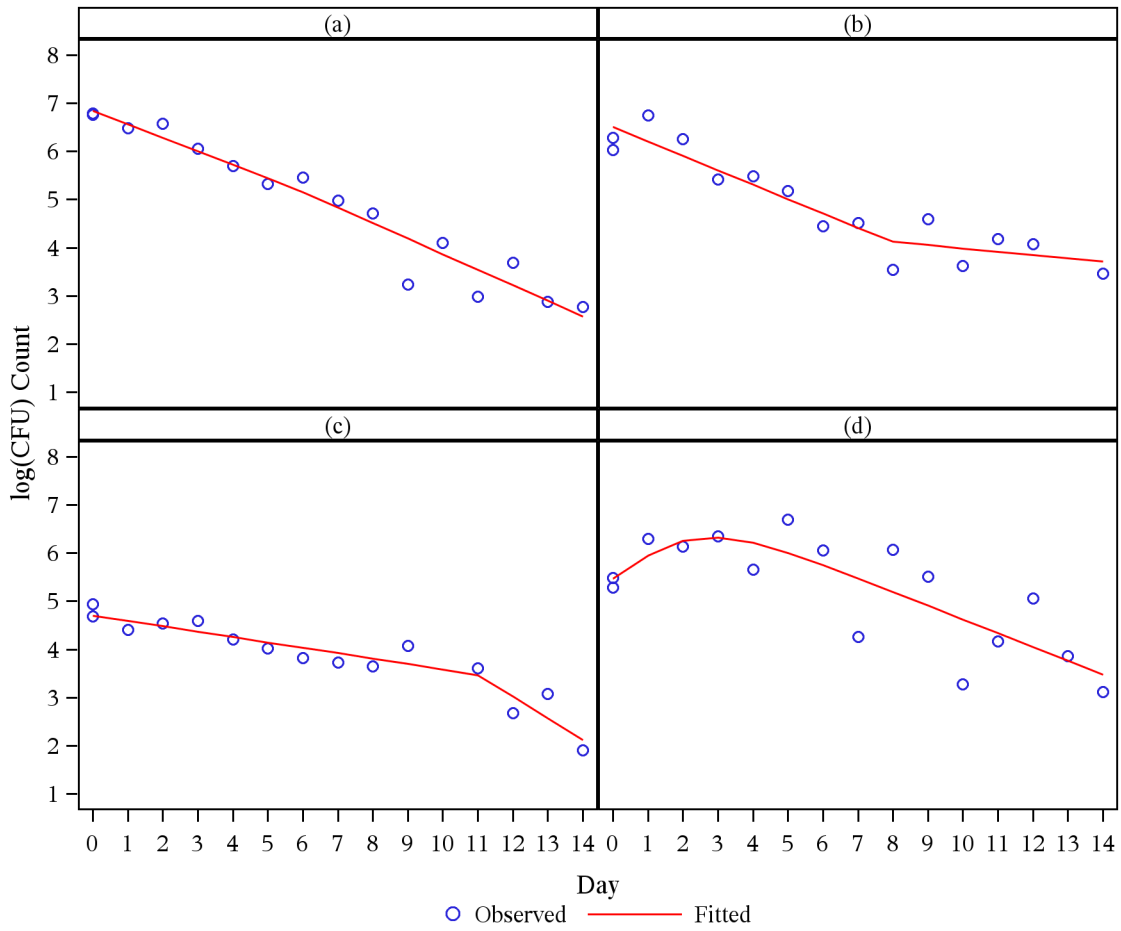**Figure 6.1:** Fitted log(CFU) Counts Versus Time for Empirical Study

Table 6.2: "By-Patient" Regression Model Parameter Estimates for Empirical Study

| Clinical Trial | Treatment Group | n | Mean (Range) | | | n_L | n_B | n_{BFS} | n_{BSF} | n_{BI} | n_{BM} |
			κ	λ_1	λ_2						
CL001	TMC207 100 mg	12	5.8 (2.0-11.0)	0.082 (-0.276-0.782)	0.062 (-0.093-0.303)	2	10	5	5	9	1
	TMC207 200 mg	13	6.6 (2.0-10.8)	-0.054 (-0.385-0.099)	0.167 (-0.006-0.497)	1	12	1	11	11	1
	TMC207 300 mg	13	6.3 (2.0-11.0)	0.058 (-0.070-0.446)	0.122 (-0.158-0.388)	3	10	3	7	9	1
	TMC207 400 mg	14	6.8 (2.0-11.0)	0.074 (-0.280-0.289)	0.141 (-0.066-0.463)	9	5	1	4	5	0
	Rifafour	6	3.8 (2.0-11.0)	0.283 (-0.278-0.534)	-0.110 (-0.957-0.167)	0	6	5	1	5	1
	Total	58	6.1 (2.0-11.0)	0.065 (-0.385-0.782)	0.100 (-0.957-0.497)	15	43	15	28	39	4
CL007	PA-824 200 mg	12	6.5 (2.0-11.0)	0.202 (-0.565-0.618)	-0.019 (-0.348-0.226)	0	12	9	3	11	1
	PA-824 600 mg	12	6.9 (2.8-11.0)	0.135 (-0.233-0.326)	0.045 (-0.271-0.269)	1	11	7	4	11	0
	PA-824 1000 mg	15	6.7 (2.0-11.0)	0.111 (-0.541-0.490)	0.052 (-0.231-0.539)	1	14	8	6	11	3
	PA-824 1200 mg	11	6.9 (2.0-11.0)	0.065 (-0.559-0.413)	0.056 (-0.529-0.722)	2	9	6	3	8	1
	Rifafour	7	5.0 (2.0-8.1)	0.378 (0.192-0.629)	-0.005 (-0.129-0.124)	0	7	7	0	6	1
	Total	57	6.5 (2.0-11.0)	0.159 (-0.565-0.629)	0.029 (-0.529-0.722)	4	53	37	16	47	6
CL010	PA-824 50 mg	12	6.1 (2.0-11.0)	0.141 (-0.040-0.525)	0.045 (-0.168-0.366)	3	9	7	2	9	0
	PA-824 100 mg	15	6.1 (2.0-11.0)	0.084 (-1.032-0.629)	0.049 (-0.131-0.244)	2	13	8	5	10	3
	PA-824 150 mg	14	6.5 (2.0-11.0)	0.022 (-0.439-0.302)	0.024 (-0.287-0.284)	2	12	8	4	12	0
	PA-824 200 mg	14	5.8 (2.0-10.0)	0.193 (-0.046-0.648)	0.108 (-0.022-0.419)	3	11	8	3	10	1
	Rifafour	8	6.7 (2.0-11.0)	0.370 (0.060-1.144)	0.166 (-0.033-0.397)	0	8	5	3	7	1
	Total	63	6.2 (2.0-11.0)	0.141 (-1.032-1.144)	0.070 (-0.287-0.419)	10	53	36	17	48	5

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Table 6.2: "By-Patient" Regression Model Parameter Estimates for Empirical Study

Clinical Trial	Treatment Group	n	κ	Mean (Range) λ_1	λ_2	n_L	n_B	n_{BFS}	n_{BSF}	n_{BI}	n_{BM}
NC001	J	14	6.9 (2.2-11.0)	-0.014 (-0.433-0.188)	0.216 (-0.099-0.718)	2	12	2	10	10	2
	J-Z	12	5.8 (2.0-11.0)	0.046 (-0.227-0.280)	0.156 (0.042-0.322)	4	8	2	6	7	1
	J-Pa	12	6.3 (2.0-11.0)	0.081 (-0.151-0.321)	0.049 (-0.176-0.155)	1	11	6	5	10	1
	Pa-Z	13	8.0 (2.0-11.0)	0.189 (-0.663-1.122)	0.091 (-0.131-0.451)	2	11	8	3	10	1
	Pa-Z-M	10	5.7 (2.0-11.0)	0.378 (0.080-0.721)	0.060 (-0.229-0.194)	1	9	9	0	7	2
	Rifafour	8	7.3 (2.0-11.0)	0.161 (0.025-0.273)	0.179 (-0.041-0.475)	1	7	3	4	6	1
	Total	69	6.7 (2.0-11.0)	0.128 (-0.663-1.122)	0.126 (-0.229-0.718)	11	58	30	28	50	8
Total	Total	247	6.4 (2.0-11.0)	0.124 (-1.032-1.144)	0.083 (-0.957-0.722)	40	207	118	89	184	23

Note: Treatment Group: J = TMC207, J-Z = TMC207 + Pyrazinamide, J-Pa = TMC207 + PA-824, Pa-Z = PA-824 + Pyrazinamide, Pa-Z-M = PA-824 + Pyrazinamide + Moxifloxacin, Rifafour = Rifafour e-275®. n = Number of randomised patients with complete profiles. n_L = Number of linearly decreasing profiles ($|\beta_2| \leq 0.05$). n_B = Number of bi-phasic profiles ($|\beta_2| > 0.05$). n_{BFS} = Number of bi-phasic profiles which initial rate of decrease is fast, followed by slower rate of decrease ($\beta_2 < -0.05$). n_{BSF} = Number of bi-phasic profiles which initial rate of decrease is slow, followed by a faster rate of decrease ($\beta_2 > 0.05$). n_{BI} = Number of bi-linear profiles with abrupt transition between the two rates of decrease ($\gamma < 1$). n_{BM} = Number of bi-phasic profiles with smooth transition between the two rates of decrease ($\gamma \geq 1$).

Observations from the empirical study suggest the following:

- Bi-linear regression models seem adequate for the log(CFU) count versus time profiles of many patients, but certainly not for all, since a substantial minority of profiles exhibit a smooth transition between phases. Whatever the case may be, it is preferable to fit a regression model that allows for a smooth transition between phases, thereby allowing one to judge the adequacy of the bi-linear regression model.

- Bi-linear regression models need to accommodate individual variation in the node, and should estimate the node parameter from the data, rather than determining it through visual inspection.

- Bi-exponential regression models are not adequate for treatments (and individual profiles) which are associated with terminal rates of decline that are faster than initial rates of decline.

- The log(CFU) count versus time profiles suggest that the residual variance is constant over the range of fitted values, i.e. the logarithm is effective as variance stabilising transformation.

On the whole, a visual inspection of the model fits suggests that the proposed regression model generally fits the data well (see Figure A.1 through Figure A.21 in the supplementary material).

6.2   Example of Application

We fitted the Bayesian NLME regression model in Equation (15) (Model 1) to the data of the NC001 trial (see Table 6.2) (Diacon et al., 2012a), and compared its fit with that of the alternative regression models (Model 2 through Model 8).

Model Selection

Model comparison statistics for the various Bayesian NLME regression models fitted are provided in Table 6.3. The model comparison statistics appear to be sensitive to the choice of the hyper parameters of the Wishart prior distributions ('default' versus 'frequentist'): This, however, is a well know drawback (Lindley, 1993) with the use of Bayes factors. The DIC favours bi-linear models slightly over bi-phasic models, followed by linear models. The marginal likelihood (Bayes factor) criterion favours linear models, followed by bi-phasic and bi-linear models. Both the DIC and marginal likelihood (Bayes factor) criteria favour models with Student t distributed errors over those with normally distributed errors.

The ICPOs suggest all models fit the data reasonably well.

Table 6.3: Comparison of Bayesian NLME Regression Models

	DIC					% ICPO $< x$		
Model	$\overline{\mathbf{D}(\theta_m, M)}$	$\mathbf{D}(\bar{\theta}_m, M)$	p_m	**DIC**(M)	$\ln(\hat{f}(y\|M))$	$x = 40$	$x = 70$	$x = 100$
Model 1	1335.00	1144.00	191.00	1526.00^2	-1365.66^4	97.57	98.87	99.11
Model 2	1360.00	1158.00	202.70	1563.00^3	-1336.71^3	97.73	98.95	99.19
Model 3	1454.00	1273.00	180.70	1635.00^5	-1382.12^7	97.98	98.62	98.95
Model 4	1476.00	1282.00	194.40	1671.00^6	-1367.23^5	97.73	98.70	99.03
Model 5	1324.00	1127.00	197.20	1521.00^1	-1376.75^6	97.57	98.87	99.19
Model 6	1445.00	1257.00	187.40	1632.00^4	-1408.10^8	97.89	98.54	98.95
Model 7	1565.00	1398.00	167.50	1733.00^7	-1236.99^1	98.54	99.11	99.19
Model 8	1644.00	1481.00	162.50	1806.00^8	-1262.32^2	98.54	98.95	99.11

Note: CPO: Conditional posterior ordinate; ICPO: Reciprocal of CPO; DIC: Deviance information crite-rion. Model 1: Bi-phasic: Student t errors and 'default' wishart priors. Model 2: Bi-phasic: Student t errors and 'frequentist' wishart priors. Model 3: Bi-phasic: Normal errors and 'default' Wishart priors. Model 4: Bi-phasic: Normal errors and 'frequentist' Wishart priors. Model 5: Bi-linear: Student t errors and 'default' Wishart priors. Model 6: Bi-linear: Normal errors and 'default' Wishart priors. Model 7: Mono-linear: Student t errors and 'default' Wishart priors. Model 8: Mono-linear: Normal errors and 'default' Wishart priors. Superscripts indicate the ranking of model comparison statistics from least favoured (1) to most favoured (8).

Early Bactericidal Activity of Study Treatments

Posterior estimates and corresponding 95% Bayesian credibility intervals (BCIs) for the mean EBA($t_1 - t_2$) of Model 1, including pairwise comparisons versus Rifafour, are presented in Table 6.4. Posterior estimates and corresponding 95% BCIs for the mean regression model parameters of Model 1 are included as supplementary material to this paper (Table B.1). Mean EBA($0-14$) was significantly different from 0 for each treatment

regimen. Treatment with Pa-Z-M had the highest bactericidal activity both over the whole 14-day treatment period, and over the time intervals Day 0 to Day 2 and Day 2 to Day 14. These results can be compared to those published by Diacon et al. (2012a).

Posterior estimates and corresponding 95% BCIs for the mean log(CFU) count versus time profiles of the six treatment regimens are presented for Model 1 in Figure 6.2a, and for Model 2 through Model 8 as supplementary material to this paper (Figure B.1 to Figure B.7, respectively). The posterior estimates and corresponding 95% BCIs for the mean log(CFU) count versus time profiles were similar for Model 1 to Model 8.

The posterior predictive distributions of the β_{2j} (i.e. β_{2fj}) based on Model 1 are presented in Figure 6.2b for each treatment group. The estimates for the mean β_2 and β_{2f} per treatment group suggest that the initial rate of decrease in CFU count for some treatment groups containing TMC207 (i.e. J and J-Z) is slow, followed by a faster rate, and *vice versa* for the treatment groups not containing TMC207 (Pa-Z and Pa-Z-M and Rifafour). The decrease in mean log(CFU) count of J-Pa is effectively linear over time. The estimates for the mean γ per treatment group suggest that the mean log(CFU) count switches from one rate of decrease to another smoothly.

Table 6.4: Model 1 – Inferential Statistics for Mean EBA$(t_1 - t_2)$

Parameter	Treatment	n	Mean Estimate	Mean 95% BCI	Mean Versus Rifafour Estimate	Mean Versus Rifafour 95% BCI
$\textbf{EBA}_j(\textbf{0} - \textbf{14})$	J (N=15)	15	0.074	[0.010; 0.145]	-0.073	[-0.185; 0.042]
	J-Z (N=15)	15	0.133	[0.065; 0.204]	-0.013	[-0.128; 0.101]
	J-Pa (N=15)	15	0.101	[0.056; 0.146]	-0.045	[-0.147; 0.055]
	Pa-Z (N=15)	15	0.154	[0.100; 0.207]	0.007	[-0.098; 0.113]
	Pa-Z-M (N=15)	15	0.248	[0.087; 0.430]	0.102	[-0.082; 0.304]
	Rifafour (N=10)	10	0.146	[0.055; 0.238]		
$\textbf{EBA}_j(\textbf{0} - \textbf{2})$	J (N=15)	15	-0.002	[-0.086; 0.084]	-0.156	[-0.316; 0.000]
	J-Z (N=15)	15	0.069	[-0.038; 0.170]	-0.085	[-0.254; 0.081]
	J-Pa (N=15)	15	0.105	[0.019; 0.187]	-0.049	[-0.210; 0.105]
	Pa-Z (N=15)	15	0.179	[0.079; 0.277]	0.025	[-0.142; 0.187]
	Pa-Z-M (N=15)	15	0.313	[0.164; 0.460]	0.159	[-0.040; 0.355]
	Rifafour (N=10)	10	0.154	[0.021; 0.290]		
$\textbf{EBA}_j(\textbf{2} - \textbf{14})$	J (N=15)	15	0.086	[0.019; 0.170]	-0.059	[-0.185; 0.075]
	J-Z (N=15)	15	0.144	[0.066; 0.229]	-0.001	[-0.132; 0.133]
	J-Pa (N=15)	15	0.100	[0.053; 0.148]	-0.044	[-0.160; 0.072]
	Pa-Z (N=15)	15	0.149	[0.093; 0.203]	0.004	[-0.114; 0.124]
	Pa-Z-M (N=15)	15	0.238	[0.046; 0.455]	0.093	[-0.124; 0.330]
	Rifafour (N=10)	10	0.145	[0.037; 0.251]		

Note: Treatment Group: J = TMC207, J-Z = TMC207 + Pyrazinamide, J-Pa = TMC207 + PA-824, Pa-Z = PA-824 + Pyrazinamide, Pa-Z-M = PA-824 + Pyrazinamide + Moxifloxacin, Rifafour = Rifafour e-275®. EBA$(t_1 - t_2)$: Early bactericidal activity over Day t_1 to Day t_2; BCI: Bayesian credibility interval. n = Number of patients in each category.

The OpenBUGS code for the implementation of Model 1 is included as supplementary material to this paper (Section C).

Figure 6.2: Model 1 – Mean log(CFU) Count and Posterior Predictive Distributions
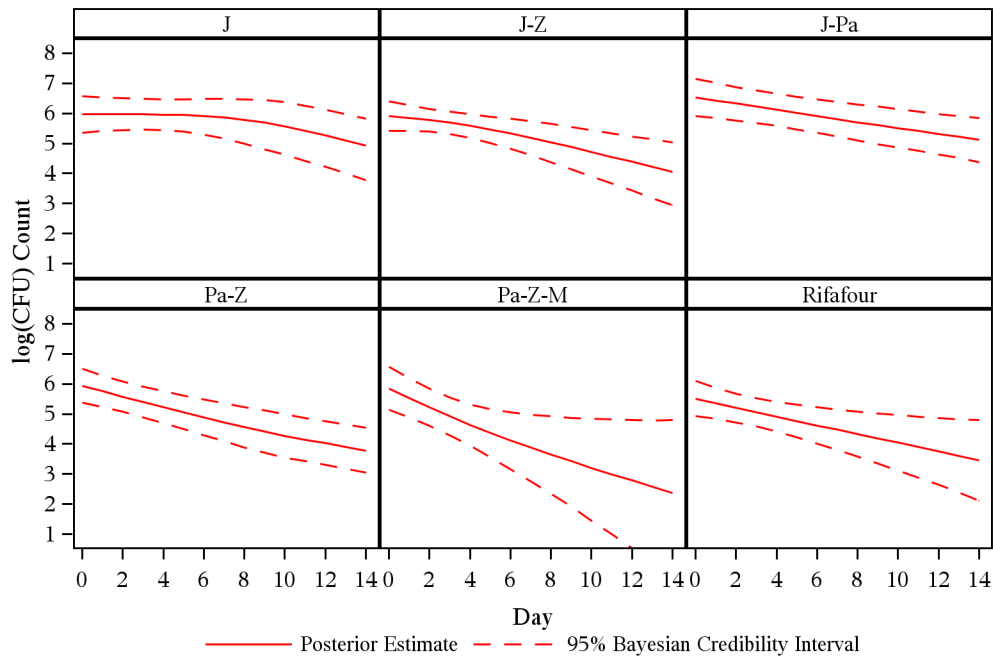
(a) Posterior Estimates and Corresponding 95% BCIs for Mean log(CFU) Count Over Time
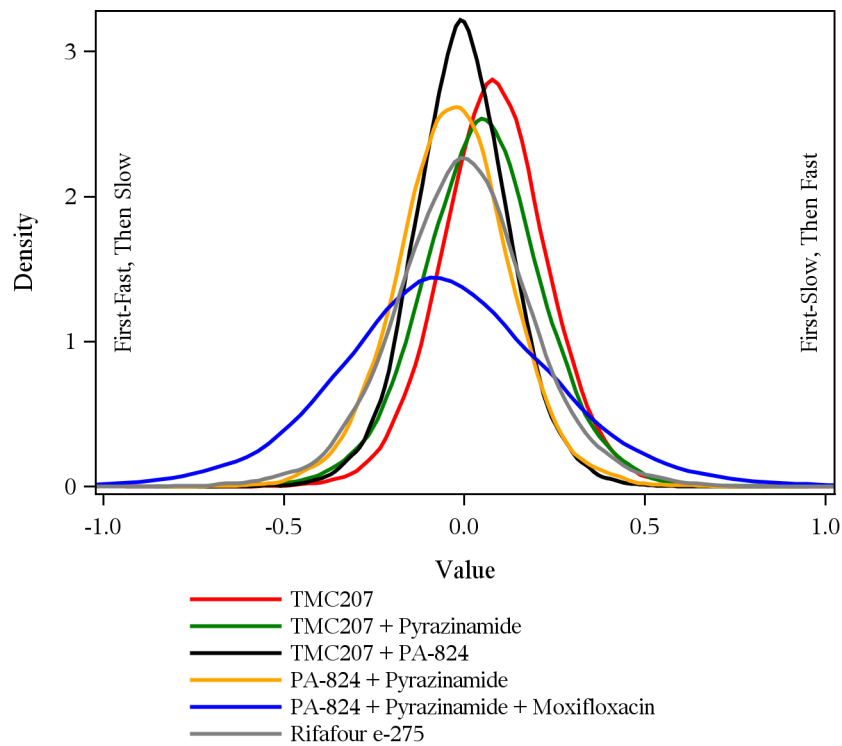
(b) Posterior Predictive Distribution of β_2 (i.e. β_{2f})

7 DISCUSSION

EBA trials of TB treatments assess the decline, during the first few days to weeks of treatment, in CFU count of *Mycobacterium tuberculosis* in the sputum of patients with smear-microscopy-positive pulmonary TB (Diacon et al., 2012a). EBA trials are a mainstay in the early clinical development of TB treatment regimens, and thus are frequently performed.

The research reported in this paper was motivated by the need for a general and flexible regression model for CFU count versus time data. Such data have conventionally been modelled using linear, bi-linear or bi-exponential regression. Linear regression, while potentially appropriate for some individual profiles, is not generally adequate since many data profiles are clearly bi-phasic, at least for treatment and observation periods longer than 2 to 7 days. Both bi-linear and bi-exponential models seem adequate for many individual profiles, but the former do not allow for a smooth transition between the initial and terminal phase of decline of CFU counts, while the latter cannot account for drugs and individual profiles which are associated with terminal rates of decline that are faster than initial rates of decline. Such terminal rates of decline have been described only recently.

In this paper, we have proposed a bi-phasic non-linear regression model for CFU data that comprises linear and bi-linear regression models as special cases, and is more flexible than bi-exponential regression models. An extensive empirical study of a large number of CFU count versus time profiles from a database of four EBA trials suggests that the proposed model fits well virtually all individual profiles. We have implemented the model as a Bayesian NLME regression model, fitted jointly to the data of all patients from a trial. One advantage of the Bayesian implementation of the model is that for patients with incomplete and sparse profiles (due to missing data), it is generally plausible

as "strength is borrowed" from the remainder of the data, which manifests as random effects estimates are shrunken towards the overall mean.

Statistical inference about the mean EBA of TB treatments is based on the Bayesian NLME regression model. The posterior predictive distribution of relevant slope parameters of the Bayesian NLME regression model provides insight into the nature of the EBA of TB treatments; specifically, the posterior predictive distribution of slope parameters allows one to judge whether treatments are associated with mono-linear or bi-linear decline of log(CFU) count, and whether log(CFU) count initially decreases fast, followed by a slower rate of decrease, or *vice versa*. In this regard, our analysis of data from the NC001 trial confirms that TMC207, somewhat unusually among anti-TB treatments, is a drug associated with a terminal rate of decline in CFU count that is faster than the initial rate of decline.

Our primary Bayesian implementation of the regression model was based on the Student t error distribution and the so-called "default" Wishart prior for the covariance matrix of the random intercept and slope parameters. However, the fit of alternative specifications of error and prior distributions was also explored. It seems that the Student t distribution, which allows for heavier tails than the normal distribution, better accommodates occasional outliers seen in the data. The DICs favour bi-linear models slightly over bi-phasic models, followed by linear models, whereas the Bayes factors favour linear models, followed by bi-phasic and bi-linear models. Given the different verdicts, it should be noted that the DIC compares models conditional on their model parameters (for which their random effects are likely to enhance model fit), whereas the Bayes factors compare models on a marginal basis. With our analysis, the Bayes factors prefer the simplest model (i.e. linear) over the more refined models (i.e. bi-phasic and bi-linear), whereas the DICs prefer the latter. Note that the linear model cannot establish to which extent the bactericidal activity between initial and later phases of treatment

differs, and investigation of this difference is a crucial aspect of EBA studies.

In summary, the bi-phasic model (Model 1) proposed here empirically fits well all individual data profiles studied and, according to the marginal likelihood (Bayes factor) criterion, is favoured over the bi-linear model. Furthermore, the bi-phasic model allows one to quantify differences in early and late rates of decline of CFU counts, which is of some importance in characterising the mode of action of anti-TB treatments.

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APPENDIX

Model checking can include the assessment of the predictive performance of the regression model using the posterior predictive distribution of replicated data $\boldsymbol{y_f}$. The goodness of fit between replicated and observed data can be assessed accordingly (Ntzoufras, 2009).

The posterior predictive distribution of $\boldsymbol{y_f}$ is given by the following expression:

$$f(\boldsymbol{y_f}|\boldsymbol{y}) = \int f(\boldsymbol{y_f}, \boldsymbol{\theta}|\boldsymbol{y})d\boldsymbol{\theta} = \int f(\boldsymbol{y_f}|\boldsymbol{\theta})f(\boldsymbol{\theta}|\boldsymbol{y})d\boldsymbol{\theta} \tag{27}$$

where $\boldsymbol{y_f}$, $\boldsymbol{y}$ and $\boldsymbol{\theta}$ represent a $r \times 1$, $n \times 1$ and $d \times 1$ vector of replicated and observed data, and model parameters, respectively.

The aforementioned approach has been criticised because of its double use of the data, and as a result, Geisser and Eddy (1979) proposed the use of the leave-one-out cross-validation predictive distribution instead, namely:

$$f(y_i|\boldsymbol{y_{[i]}}) = \int f(y_i|\boldsymbol{\theta})f(\boldsymbol{\theta}|\boldsymbol{y_{[i]}})d\boldsymbol{\theta} \tag{28}$$

where $\boldsymbol{y_{[i]}}$ represents the vector $\boldsymbol{y}$ with the ith observation (i.e. y_i) omitted.

The quantity $f(y_i|\boldsymbol{y_{[i]}})$ in Equation (28) is also known as the conditional posterior ordinate, and can be estimated by the following:

$$\widehat{\text{CPO}}_i = \left(\frac{1}{L}\sum_{l=1}^{L}\frac{1}{f(y_i|\boldsymbol{\theta}^{(l)})}\right)^{-1} \tag{29}$$

where $\boldsymbol{\theta}^{(l)}$ represents the vector of posterior MCMC samples from $\boldsymbol{\theta}$ at iteration l. The $\widehat{\text{CPO}}_i$ estimate can be interpreted as the harmonic mean of the probability distribution of y_i for each $\boldsymbol{\theta}^{(l)}$, where $l = 1, 2, \ldots, L$ following the simulation burn-in period.

A large number of small $\widehat{\text{CPO}}_i$ estimates would indicate a poor fit of the candidate

model. Such $\widehat{\text{CPO}}_i$ estimates can also be used to identify possible outliers in the data. Conversely, the reciprocal of $\widehat{\text{CPO}}_i$, or $\widehat{\text{ICPO}}_i$, can also be used to assess model fit. Estimates of $\widehat{\text{CPO}}_i > 40$ and $\widehat{\text{CPO}}_i > 70$ highlight possible or extreme outliers in the data, respectively (Ntzoufras, 2009).

SUPPLEMENTARY MATERIAL
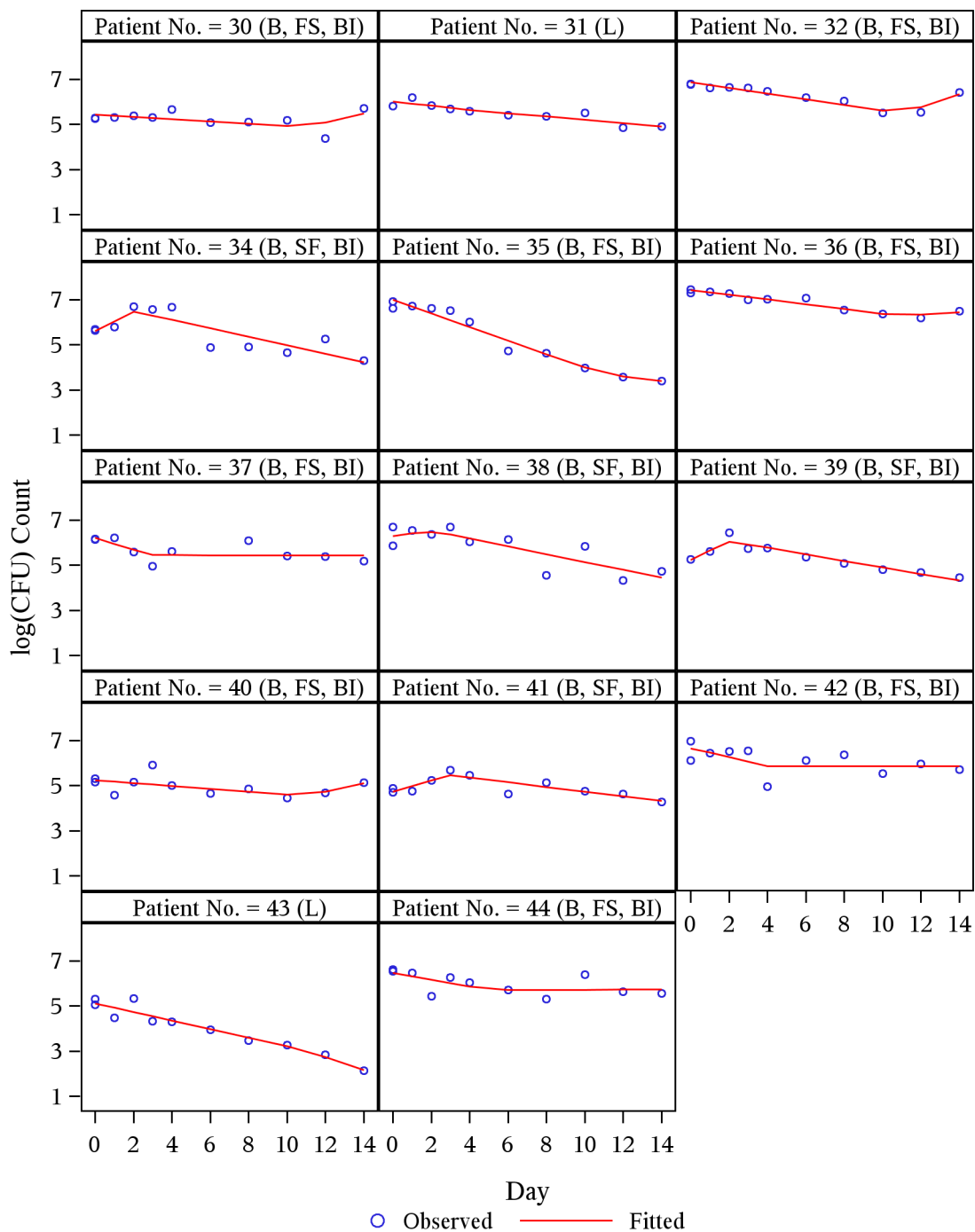
A   EMPIRICAL STUDY

Figure A.1: Observed and Fitted log(CFU) Count
Trial **CL001**, Treatment **TMC207 100 mg**
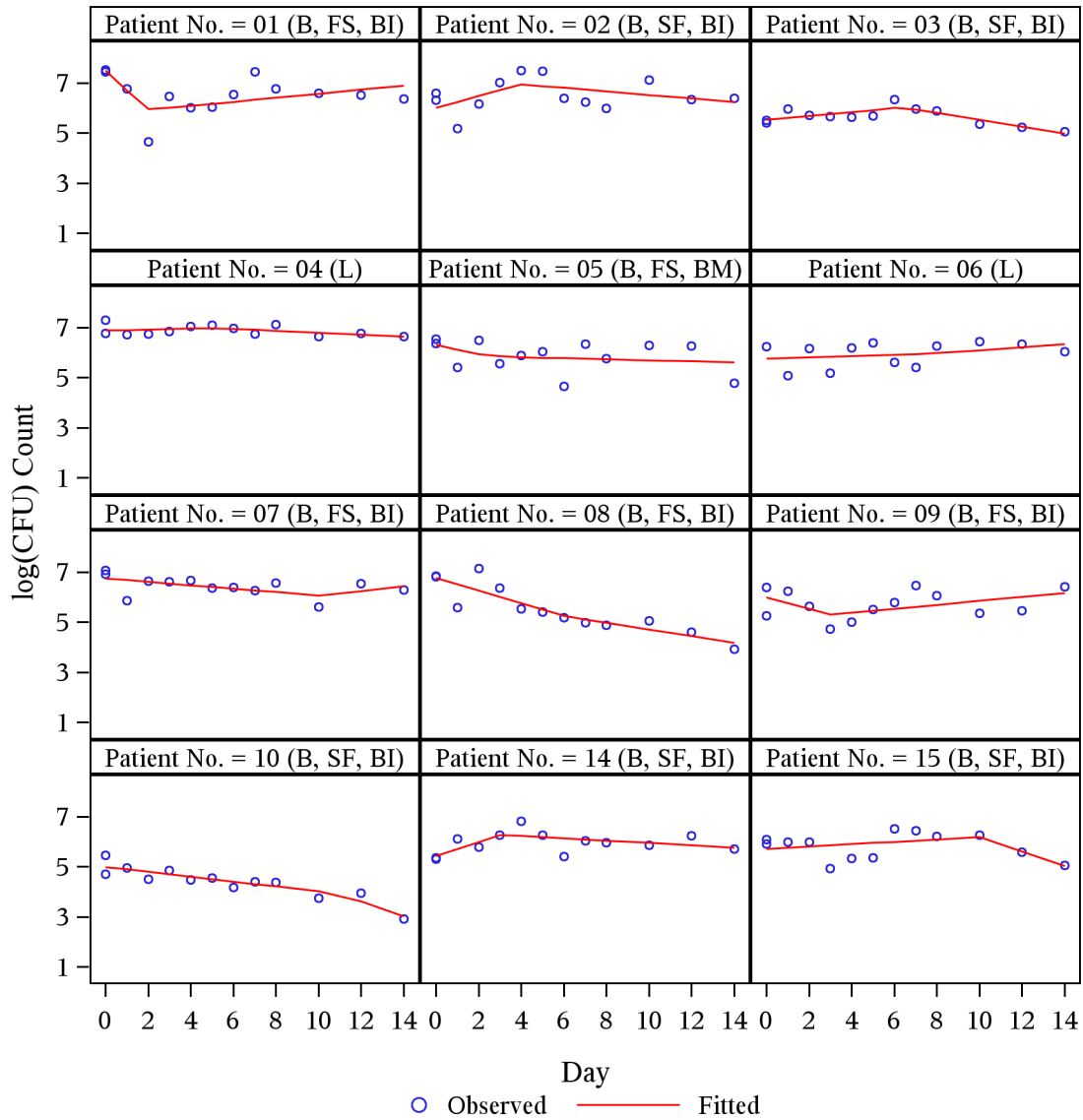
Figure A.2: Observed and Fitted log(CFU) Count
Trial **CL001**, Treatment **TMC207 200 mg**
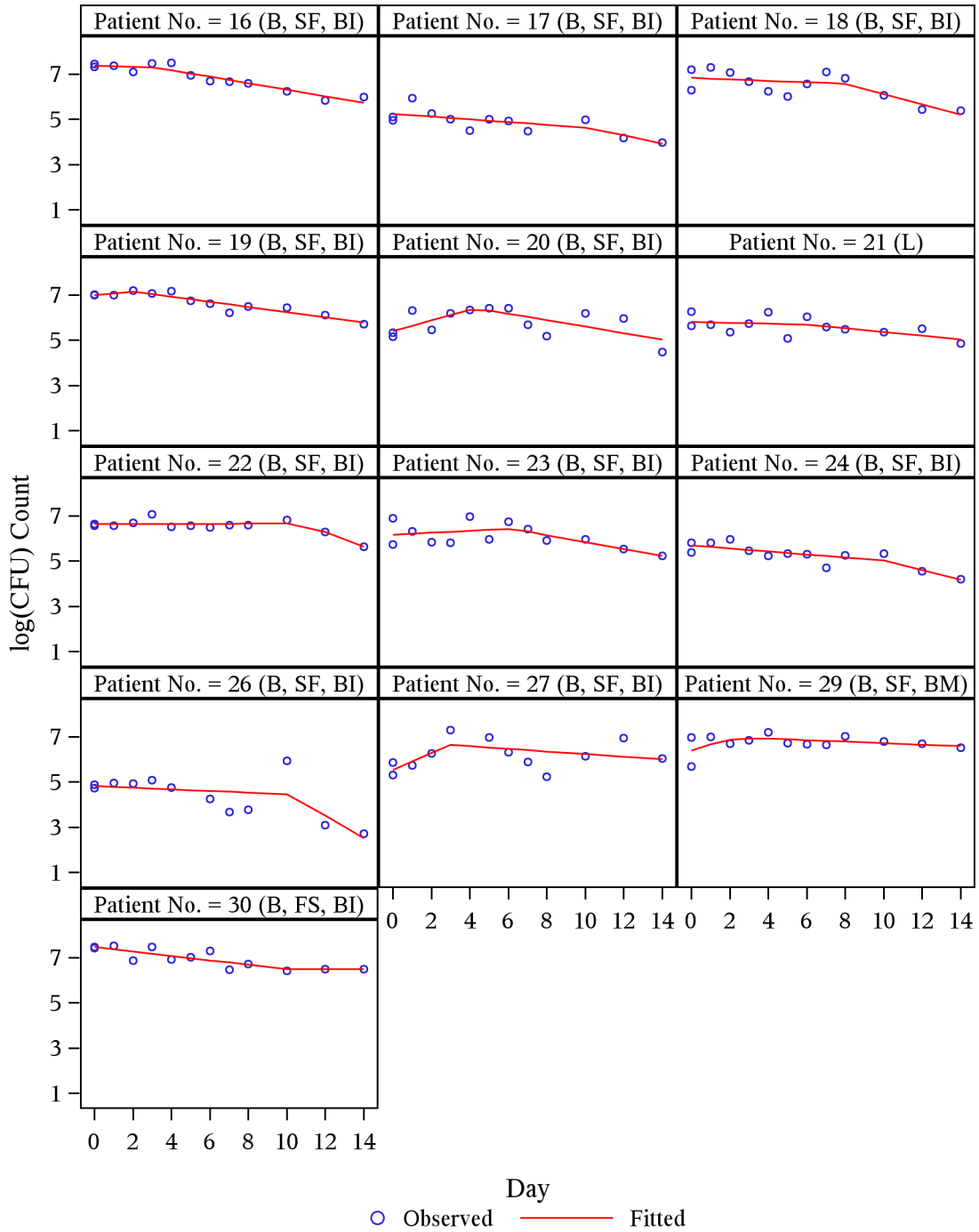
Figure A.3: Observed and Fitted log(CFU) Count
Trial **CL001**, Treatment **TMC207 300 mg**
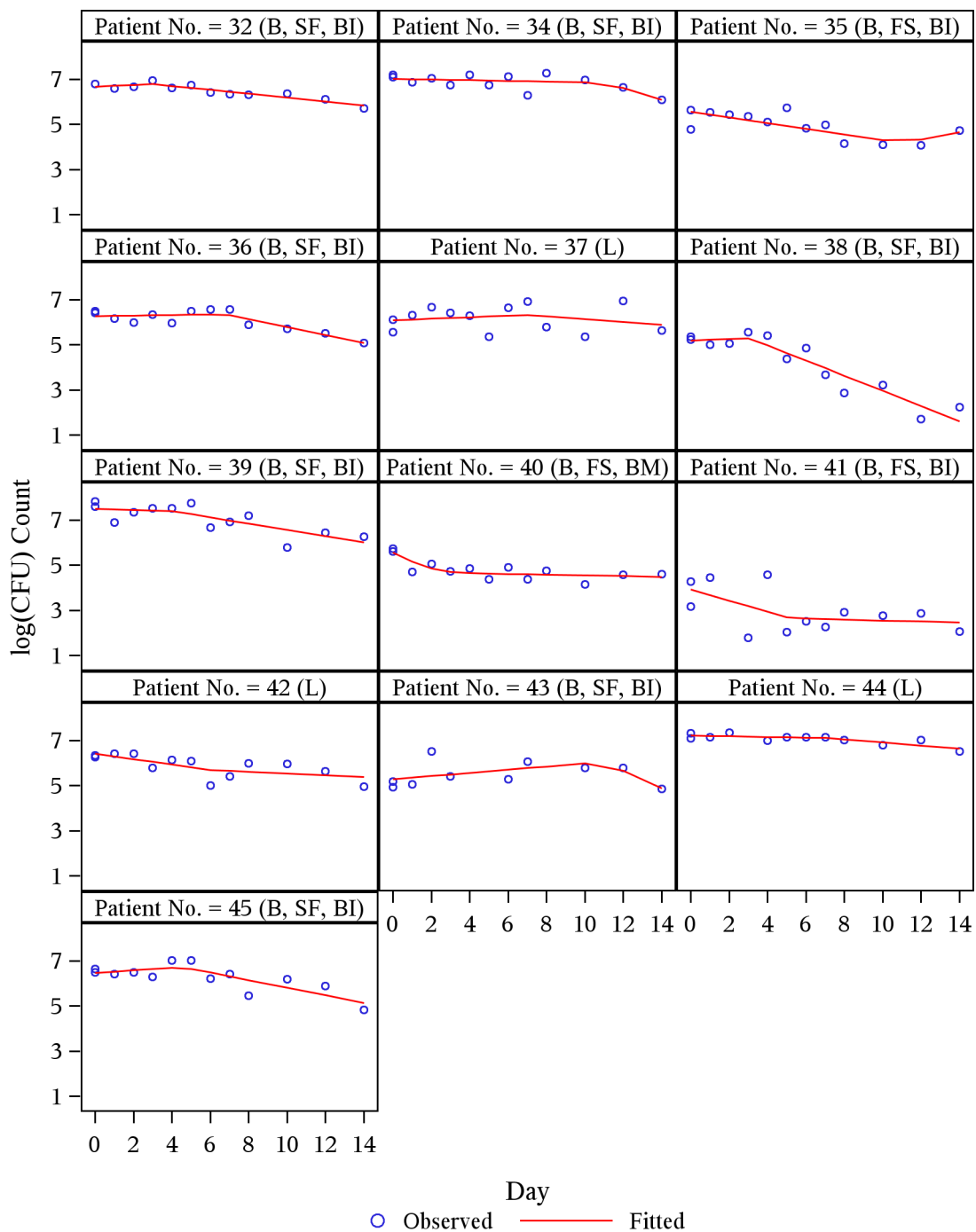
Figure A.4: Observed and Fitted log(CFU) Count
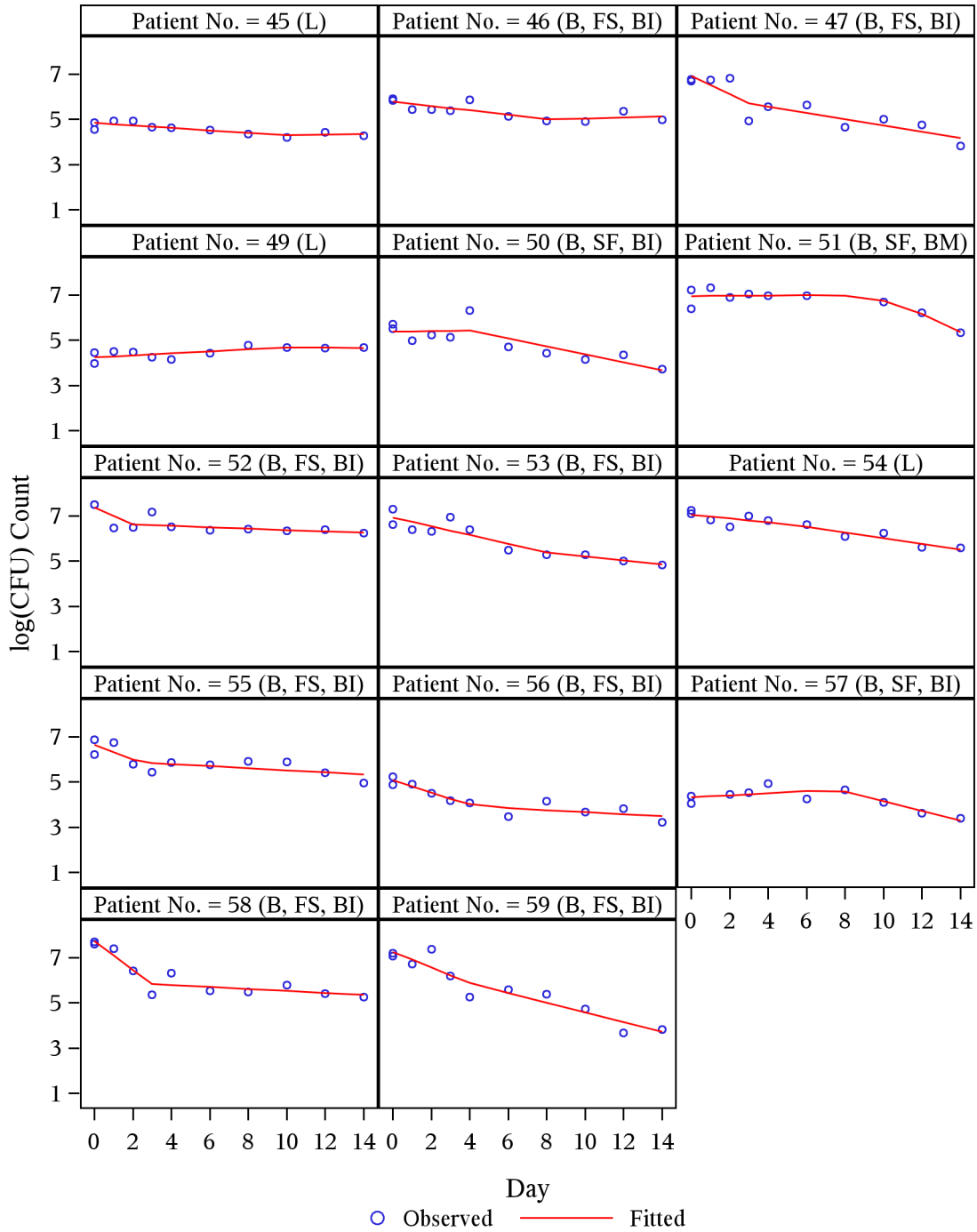Trial **CL001**, Treatment **TMC207 400 mg**
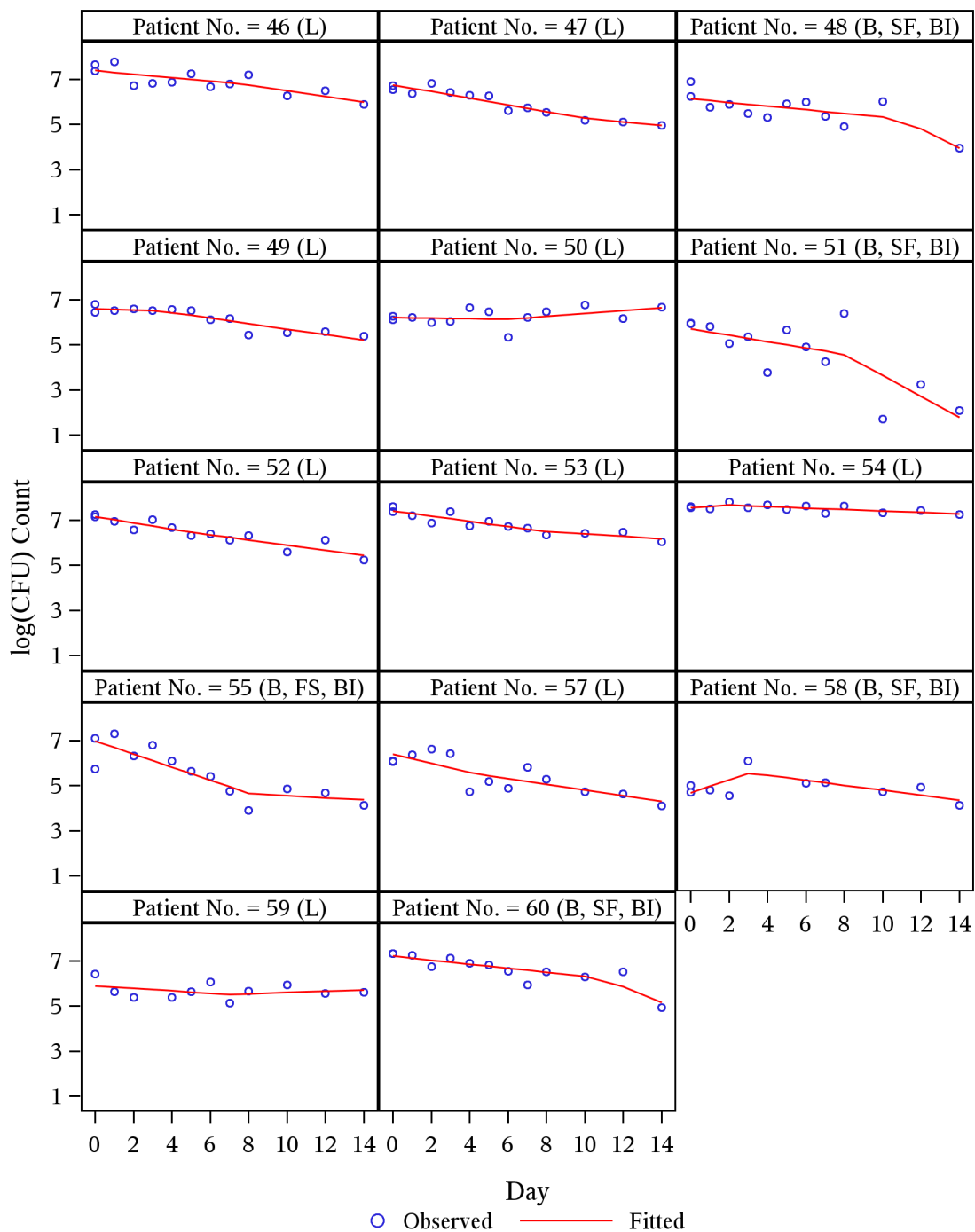
Figure A.5: Observed and Fitted log(CFU) Count
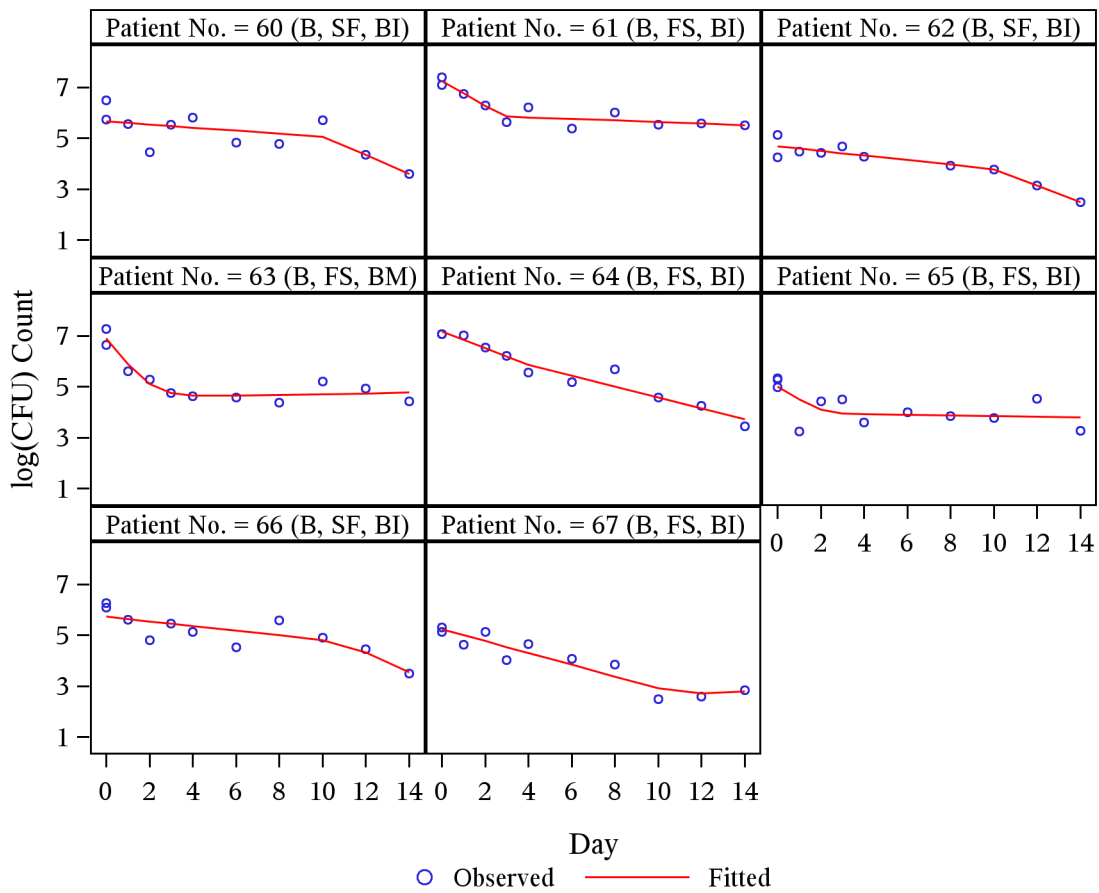Trial **CL001**, Treatment **Rifafour**
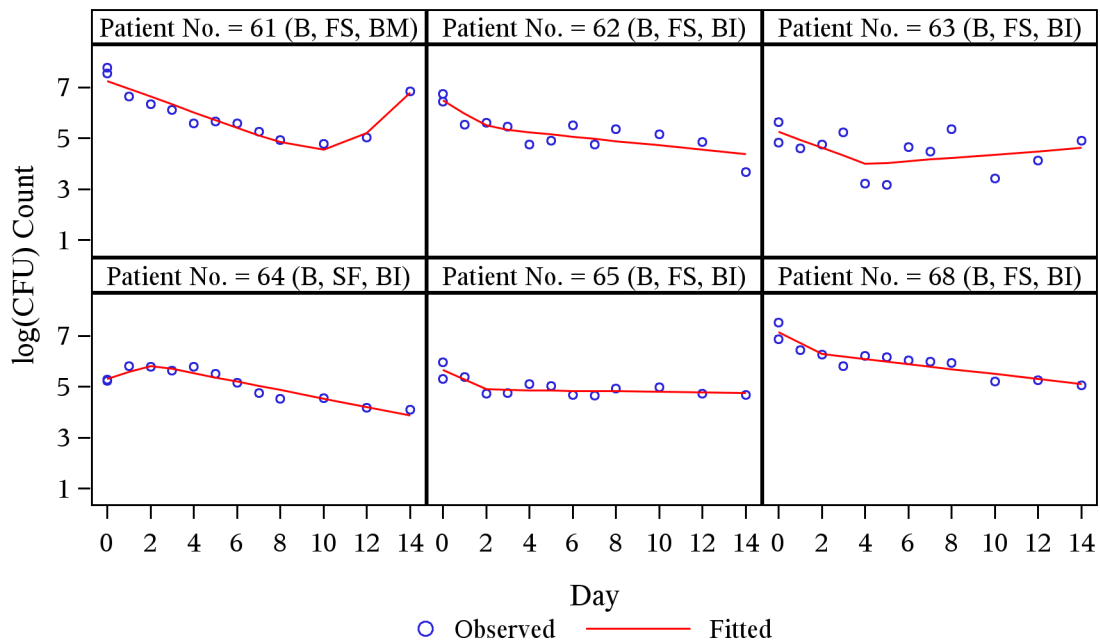
Figure A.6: Observed and Fitted log(CFU) Count
Trial **CL007**, Treatment **PA-824 200 mg**
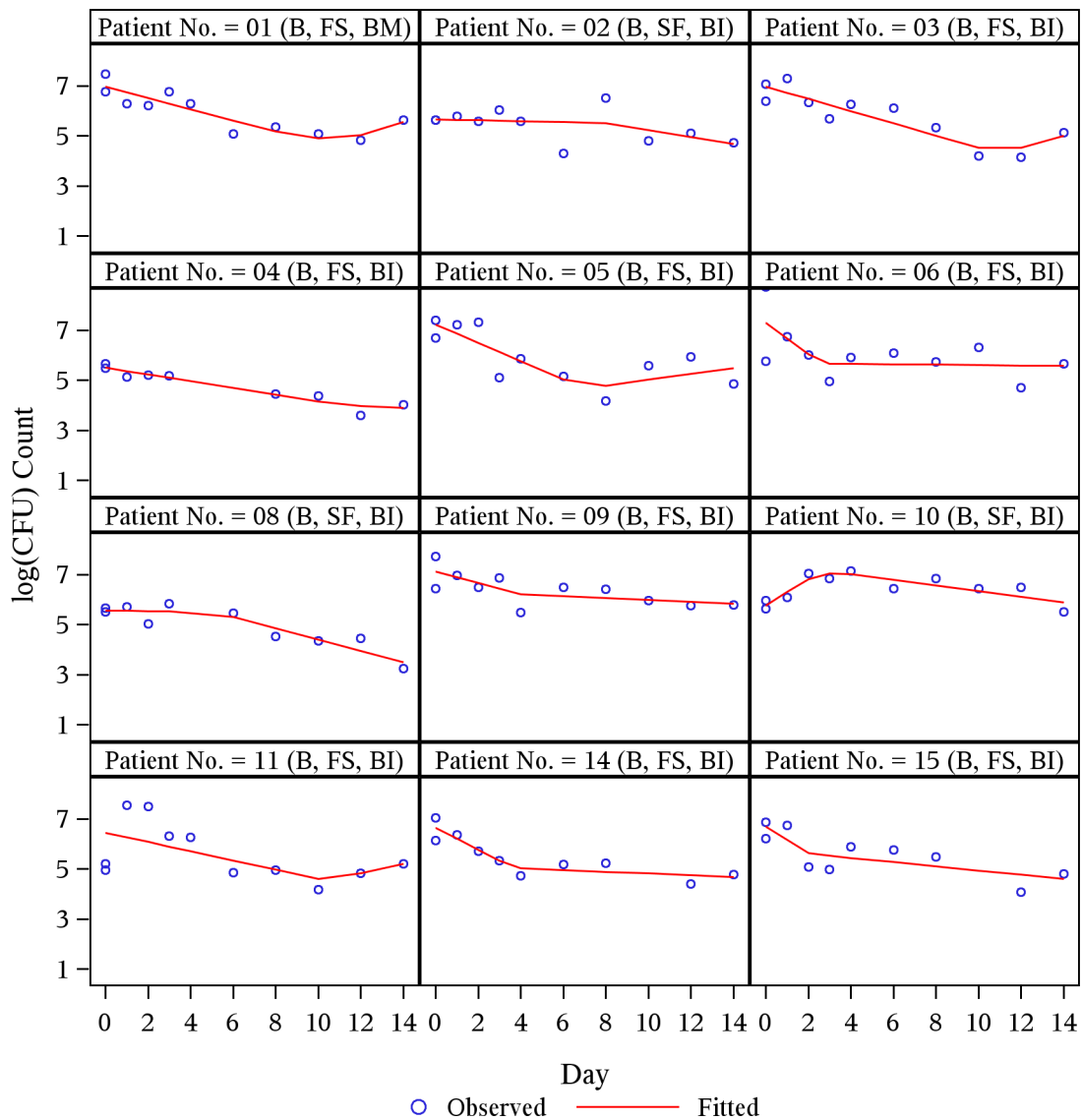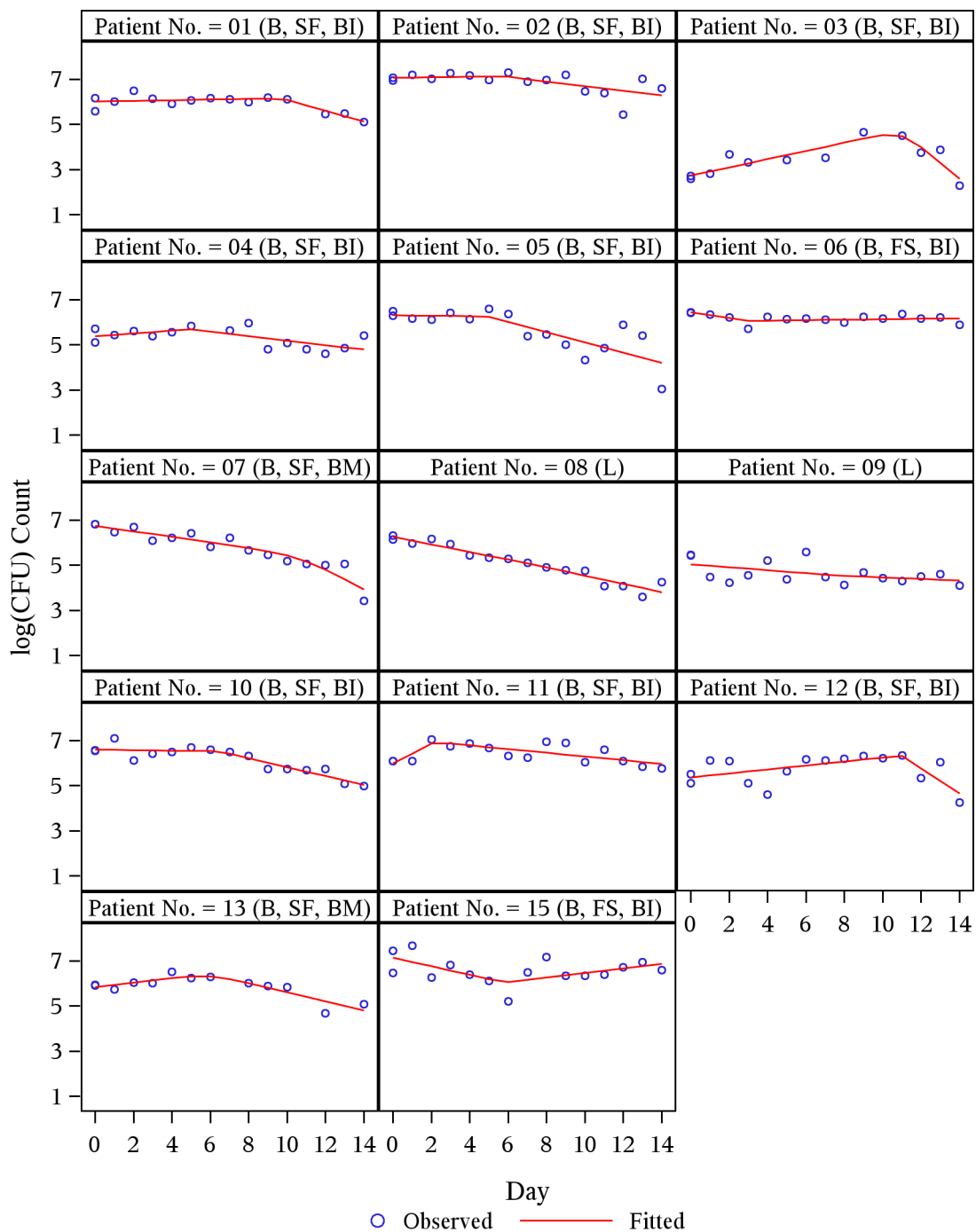
Figure A.7: Observed and Fitted log(CFU) Count
Trial **CL007**, Treatment **PA-824 600 mg**
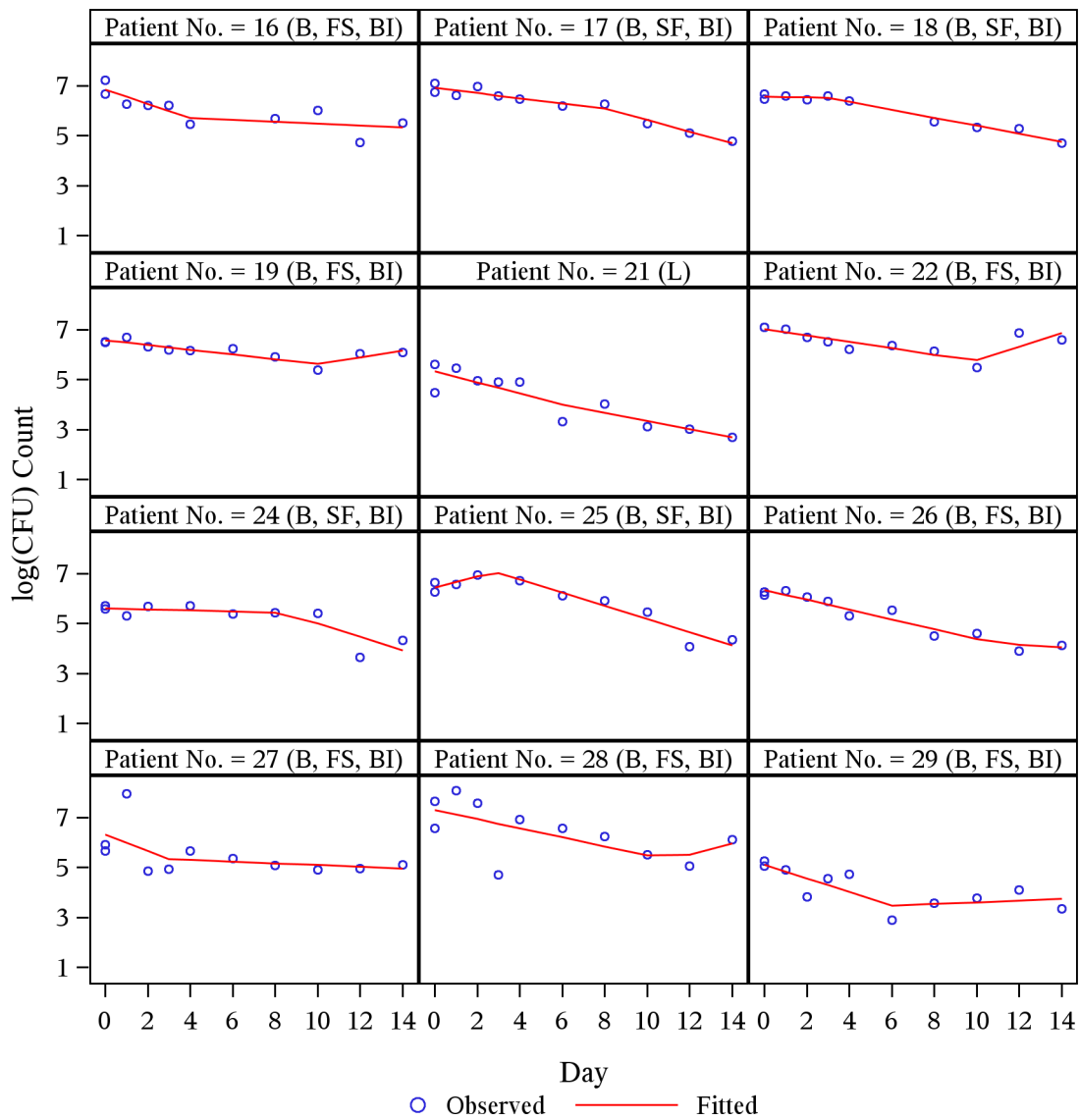
Figure A.8: Observed and Fitted log(CFU) Count
Trial **CL007**, Treatment **PA-824 1000 mg**
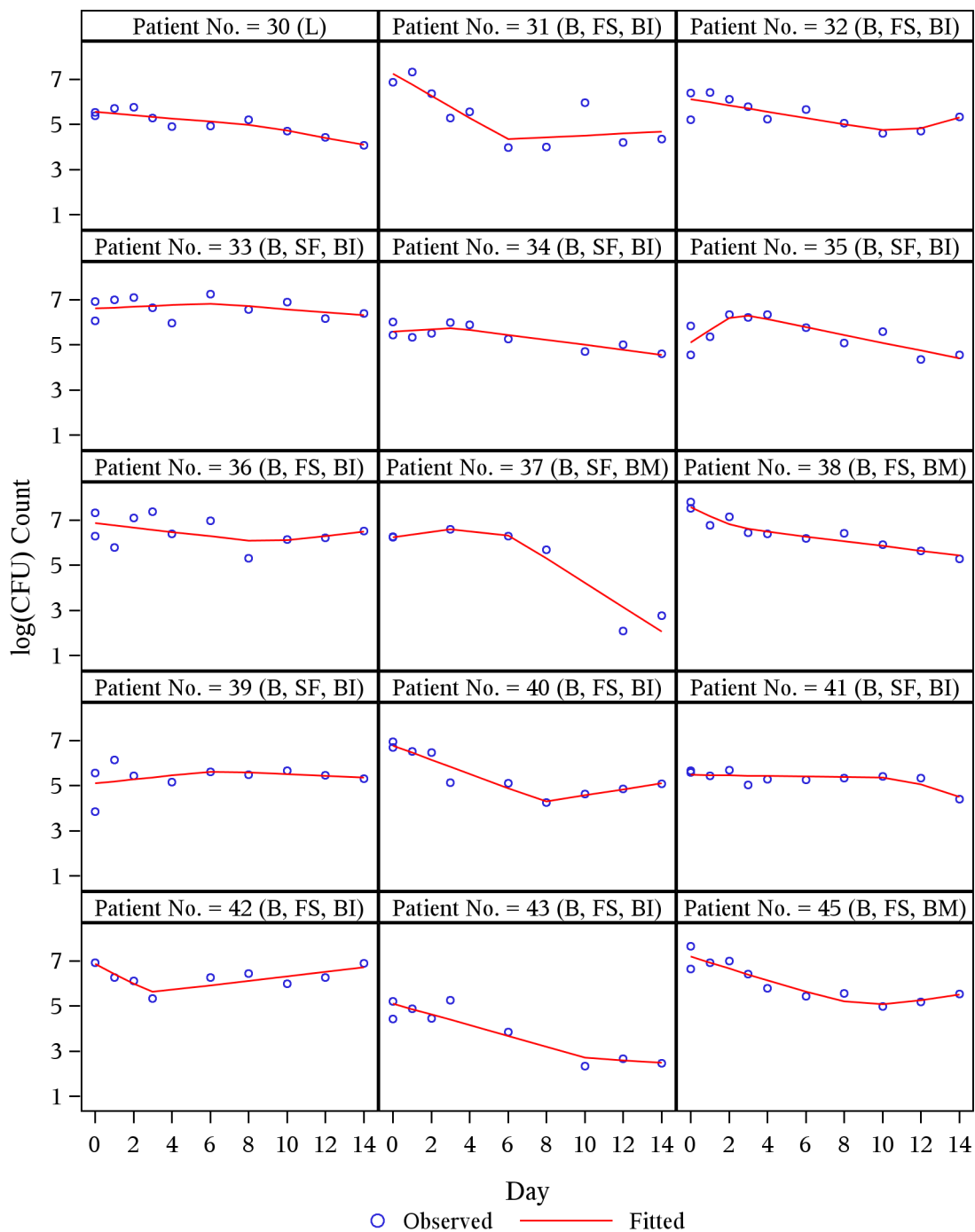
Figure A.9: Observed and Fitted log(CFU) Count
Trial **CL007**, Treatment **PA-824 1200 mg**
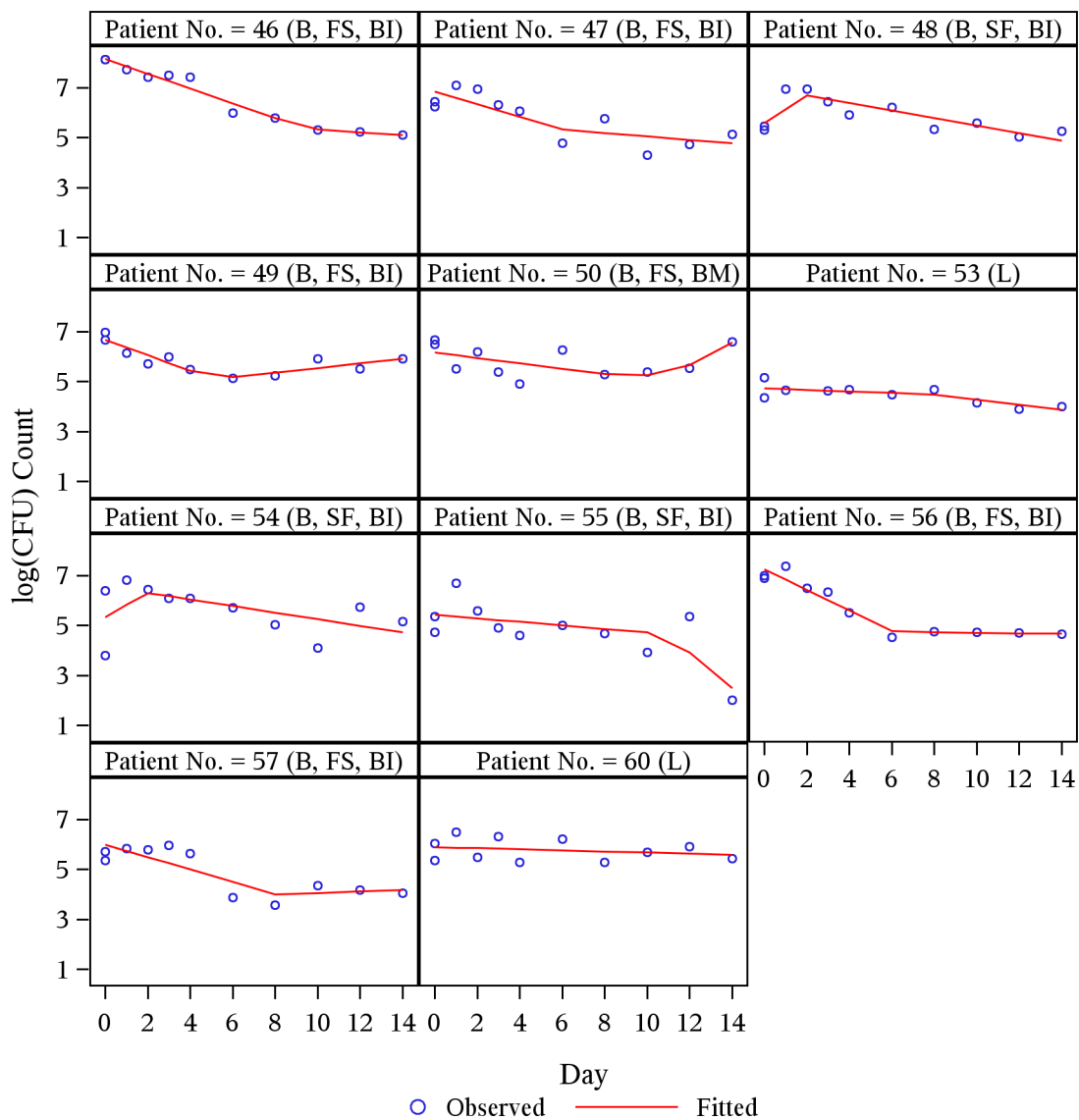
Figure A.10: Observed and Fitted log(CFU) Count
Trial **CL007**, Treatment **Rifafour**
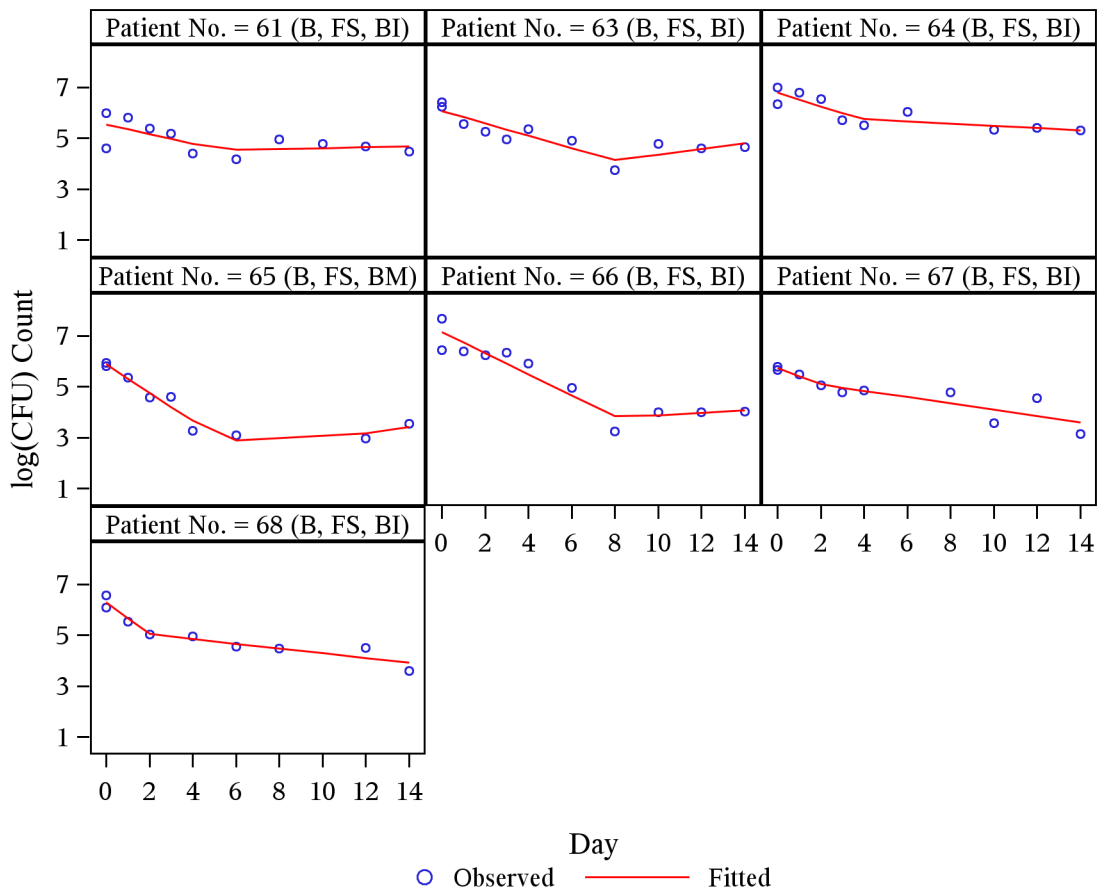
Figure A.11: Observed and Fitted log(CFU) Count
Trial **CL010**, Treatment **PA-824 50 mg**
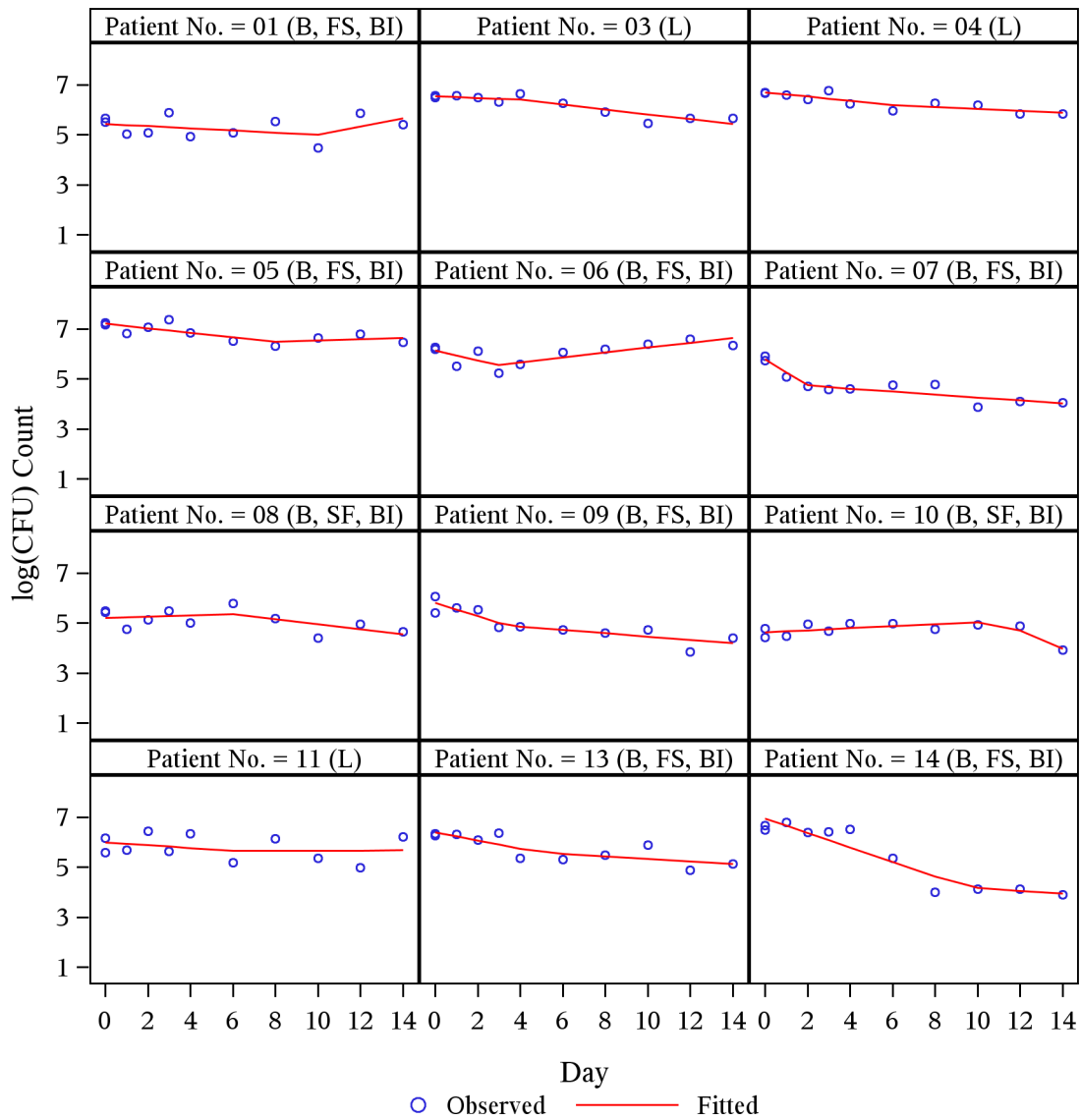
Figure A.12: Observed and Fitted log(CFU) Count
Trial **CL010**, Treatment **PA-824 100 mg**
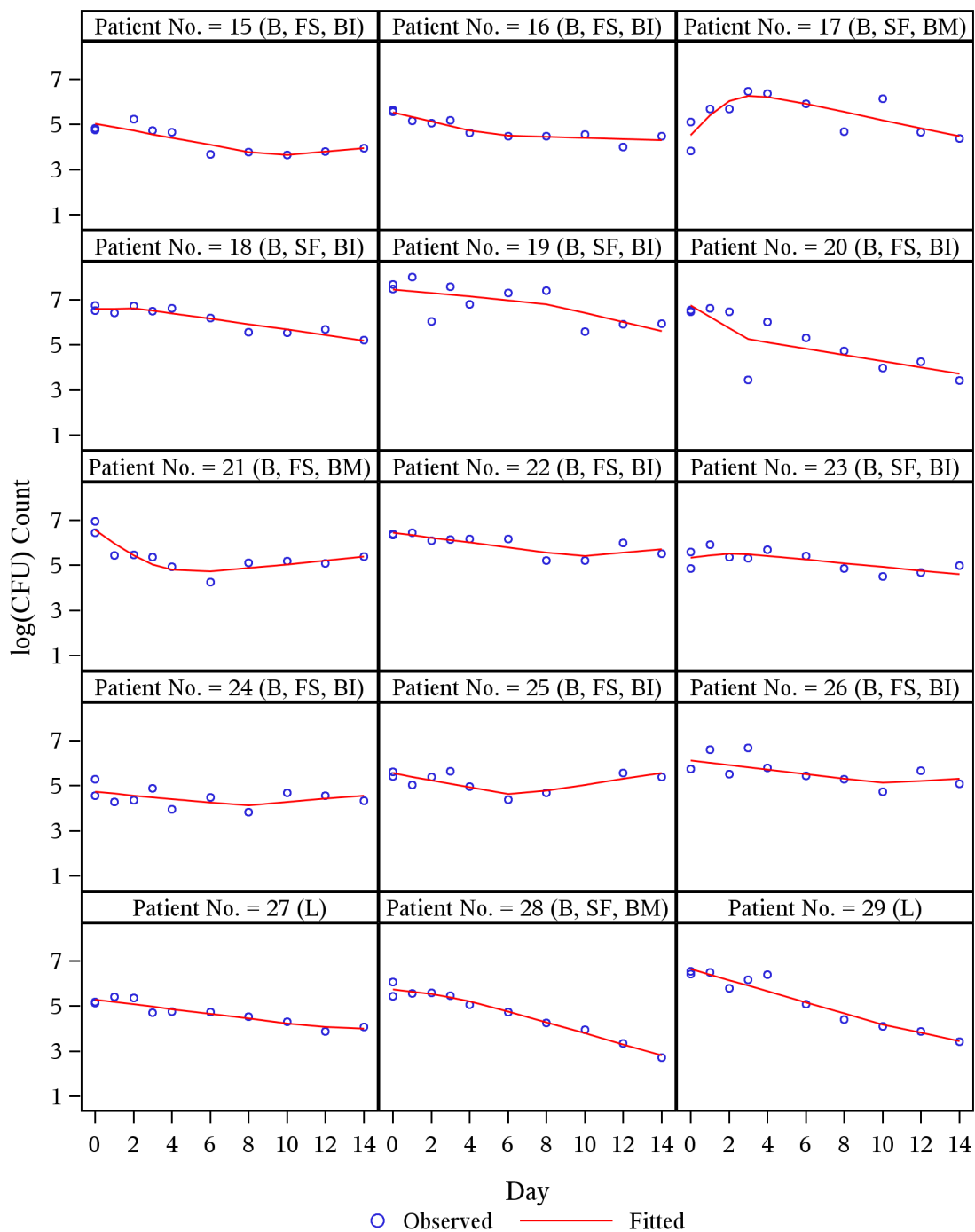
Figure A.13: Observed and Fitted log(CFU) Count
Trial **CL010**, Treatment **PA-824 150 mg**

Figure A.14: Observed and Fitted log(CFU) Count
Trial **CL010**, Treatment **PA-824 200 mg**

Figure A.15: Observed and Fitted log(CFU) Count
Trial **CL010**, Treatment **Rifafour**

Figure A.16: Observed and Fitted log(CFU) Count
Trial **NC001**, Treatment **J**

Figure A.17: Observed and Fitted log(CFU) Count
Trial **NC001**, Treatment **J-Z**
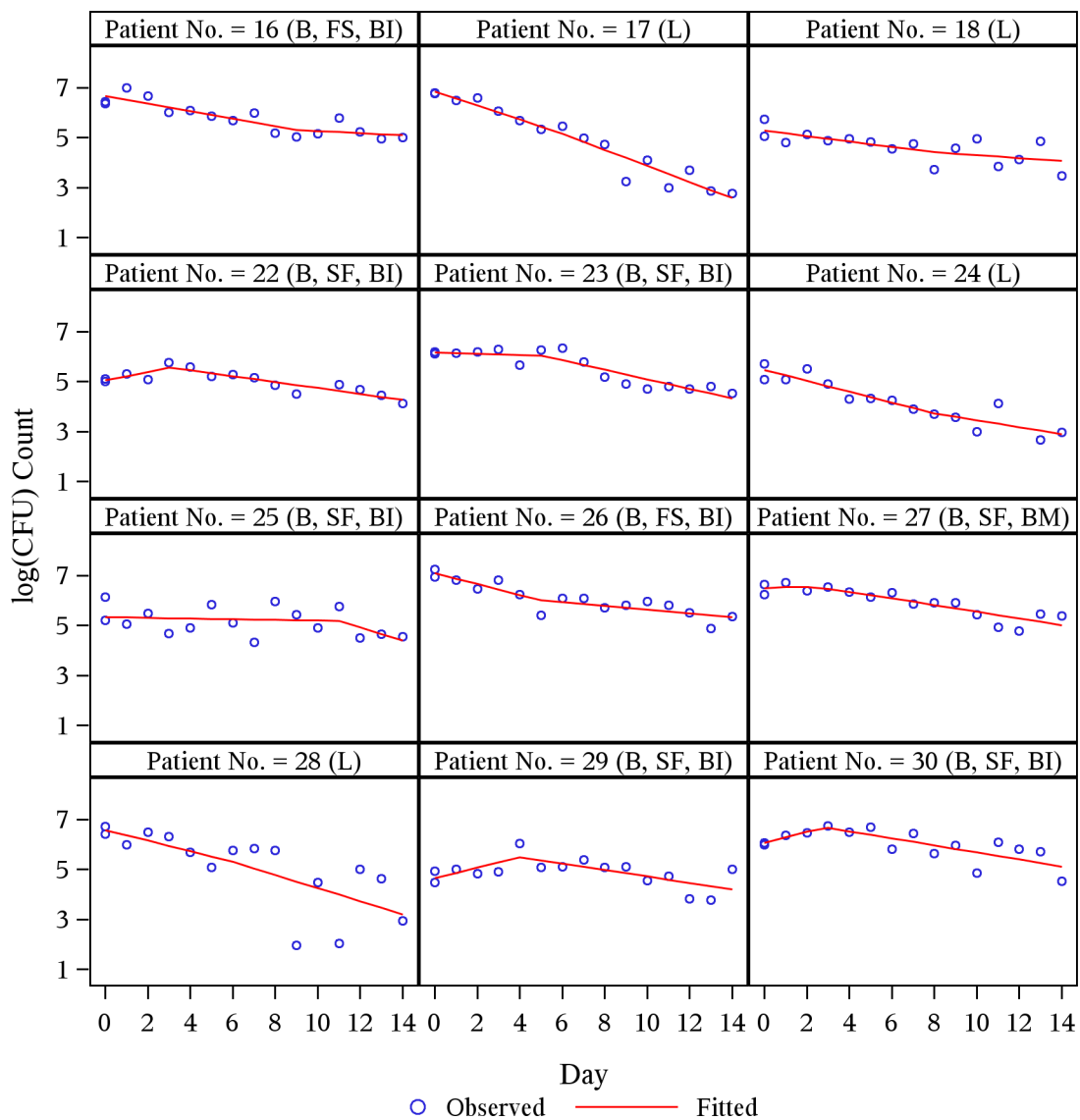
Figure A.18: Observed and Fitted log(CFU) Count
Trial **NC001**, Treatment **J-Pa**
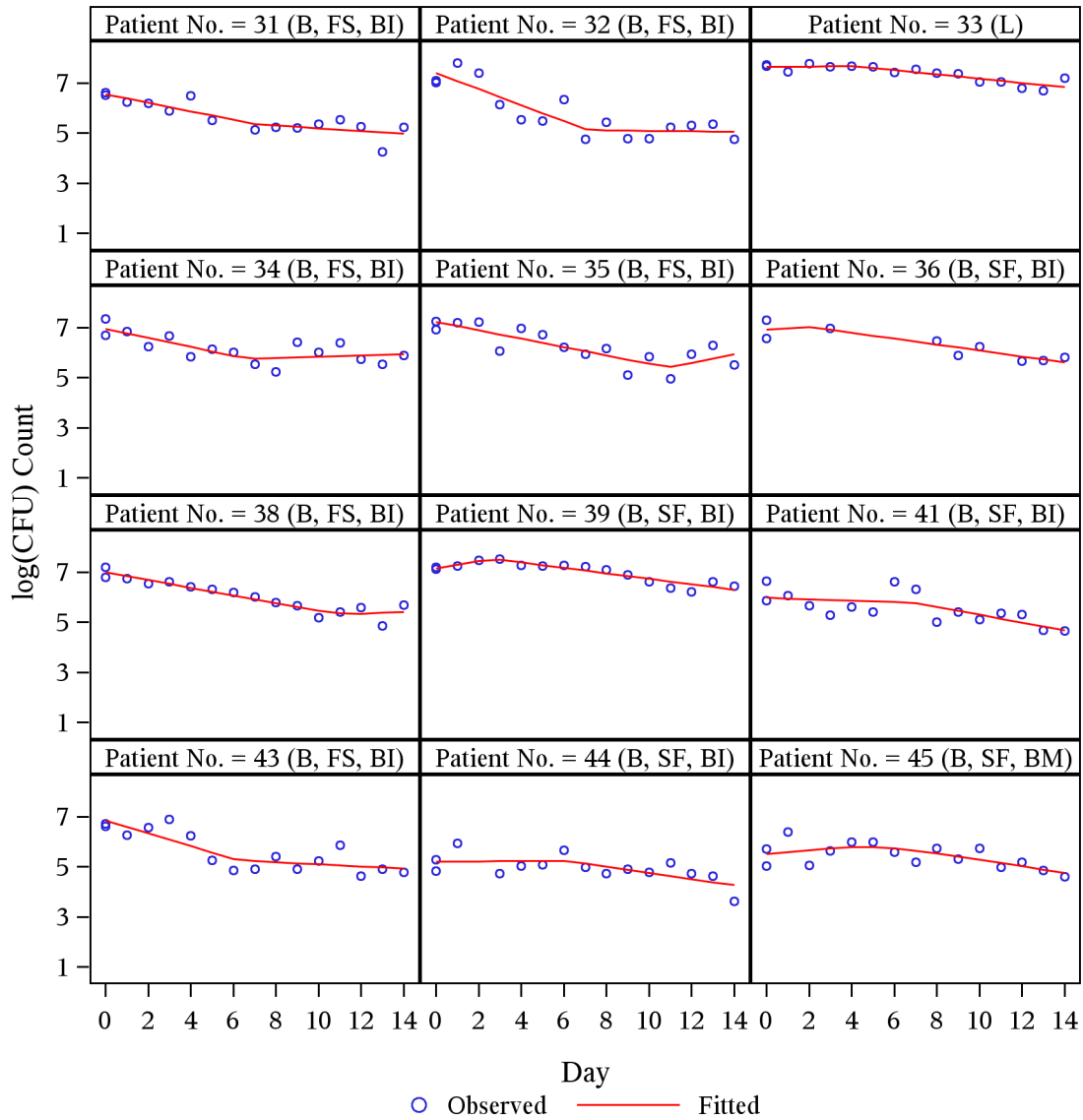
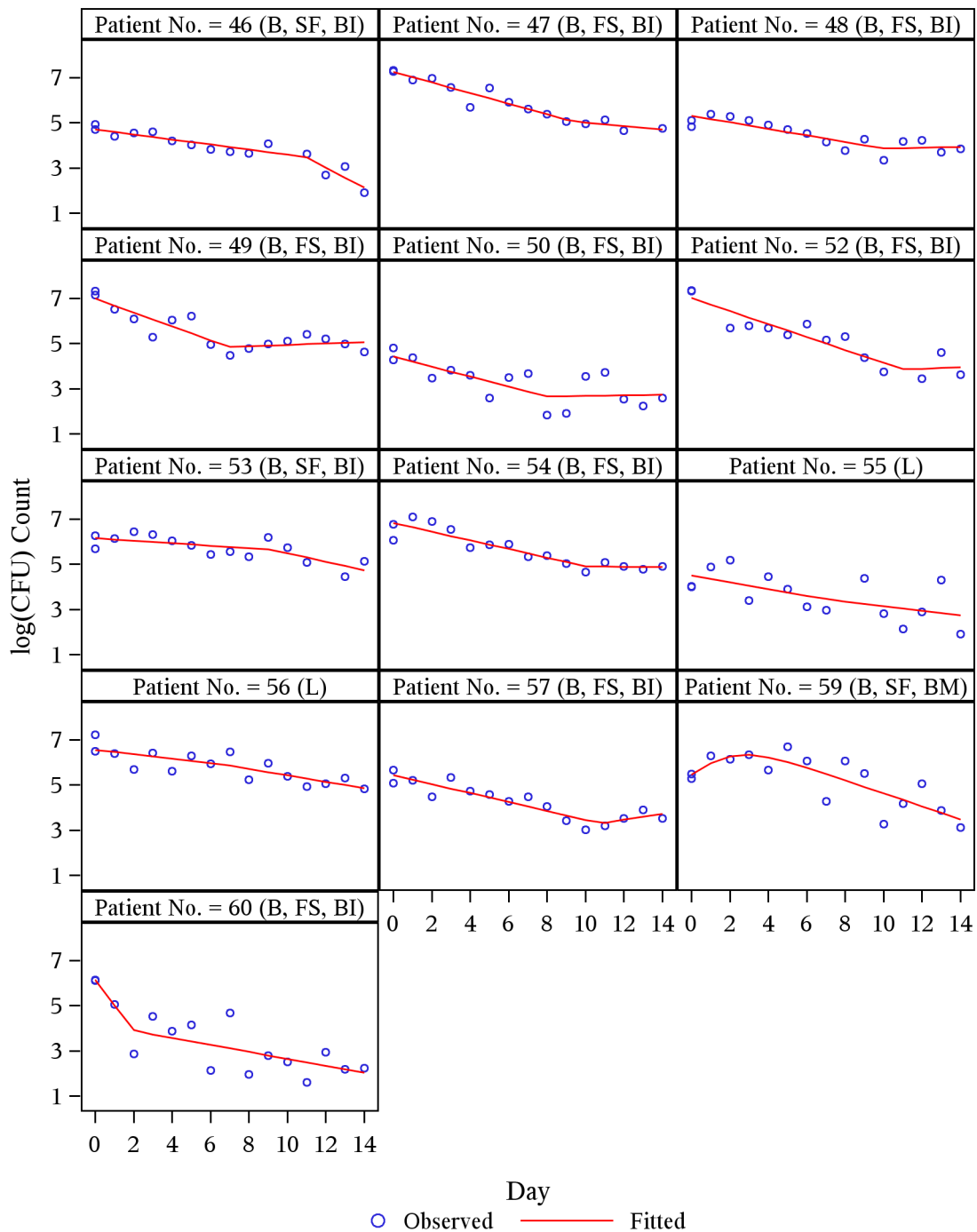**Figure A.19:** Observed and Fitted log(CFU) Count
Trial **NC001**, Treatment **Pa-Z**

Figure A.20: Observed and Fitted log(CFU) Count
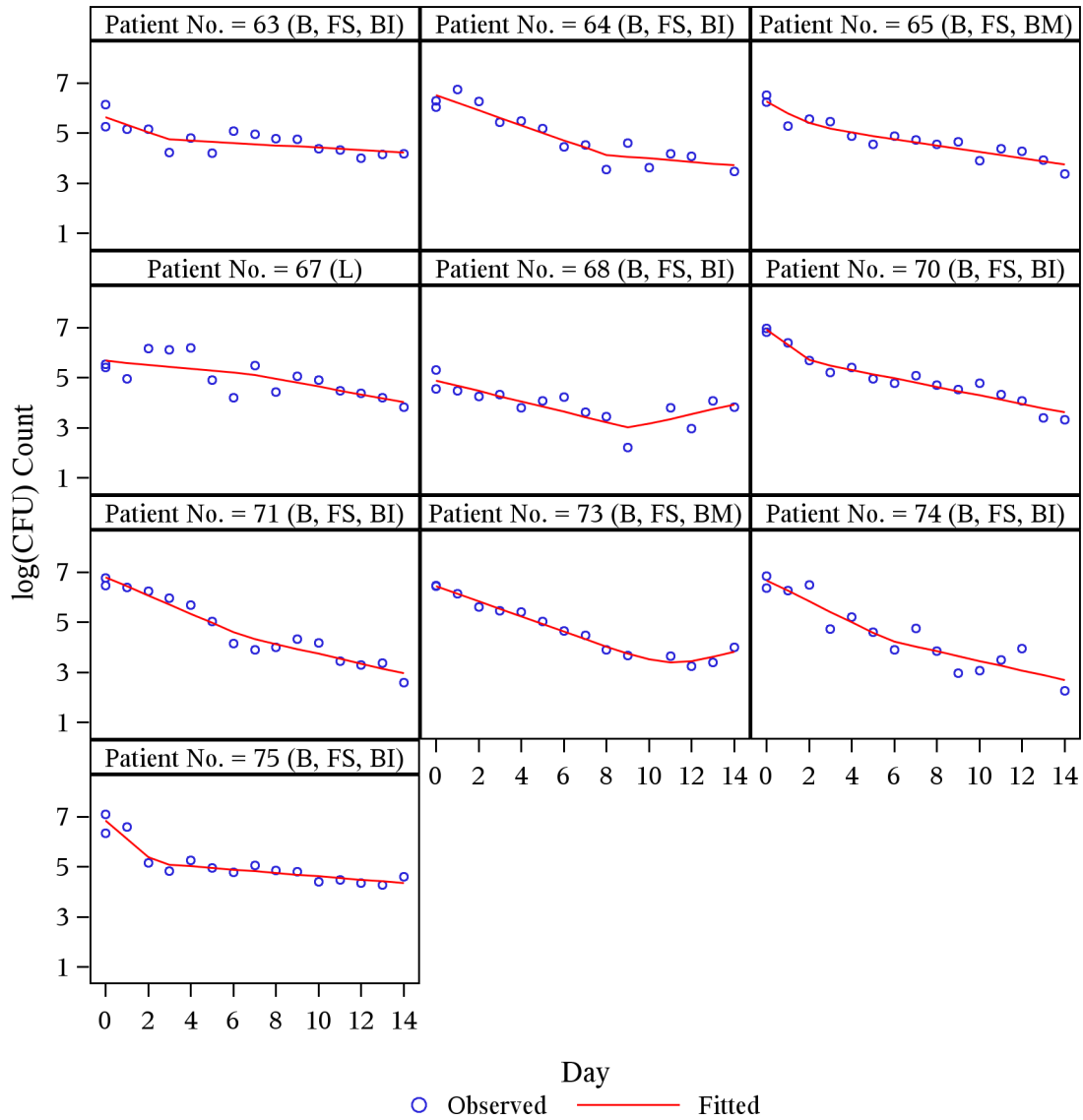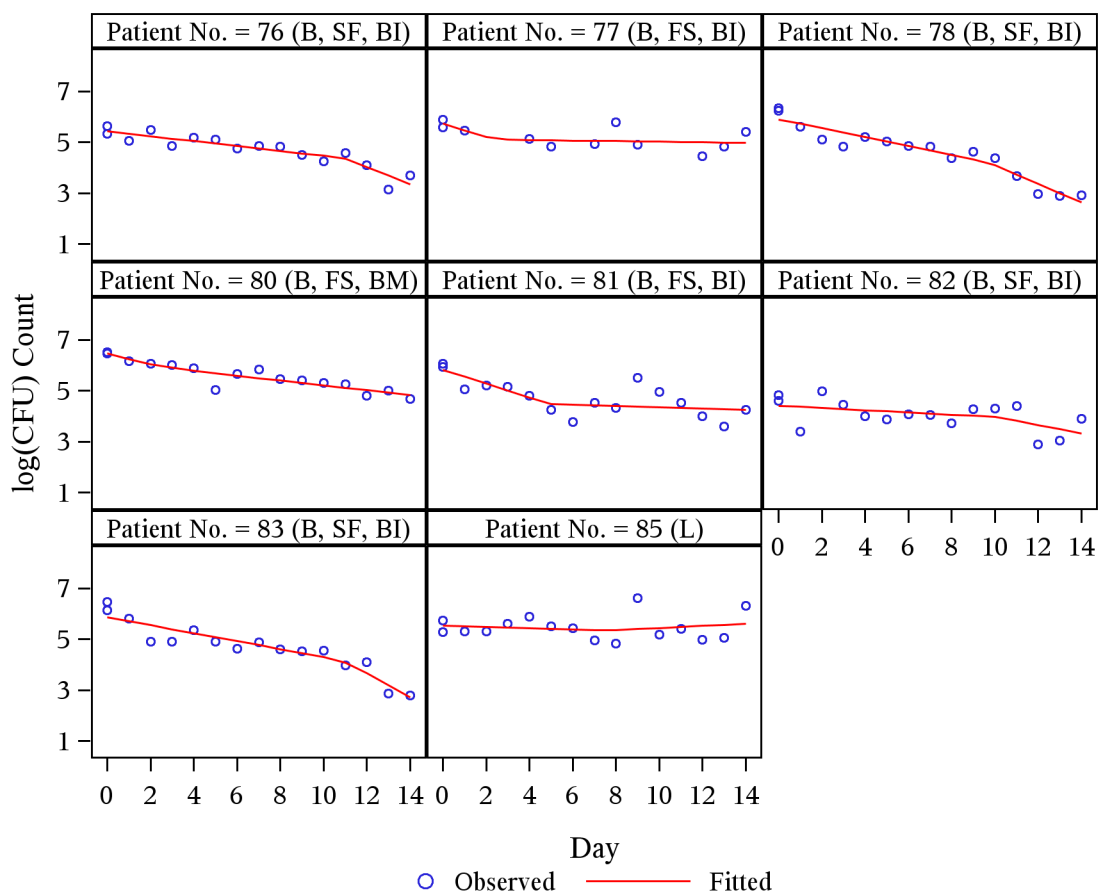Trial **NC001**, Treatment **Pa-Z-M**

Figure A.21: Observed and Fitted log(CFU) Count
Trial **NC001**, Treatment **Rifafour**

B   EXAMPLE OF APPLICATION: ADDITIONAL RESULTS

Table B.1: Model 1 – Inferential Statistics for Regression Model Parameters

Parameter	Treatment	n	Mean Estimate	95% BCI
α_j	J (N=15)	15	5.965	[5.362; 6.578]
	J-Z (N=15)	15	5.912	[5.416; 6.392]
	J-Pa (N=15)	15	6.535	[5.909; 7.152]
	Pa-Z (N=15)	15	5.934	[5.380; 6.498]
	Pa-Z-M (N=15)	15	5.844	[5.131; 6.560]
	Rifafour (N=10)	10	5.507	[4.923; 6.094]
β_{1j}	J (N=15)	15	0.081	[0.029; 0.132]
	J-Z (N=15)	15	0.116	[0.057; 0.174]
	J-Pa (N=15)	15	0.100	[0.059; 0.141]
	Pa-Z (N=15)	15	0.149	[0.101; 0.199]
	Pa-Z-M (N=15)	15	0.262	[0.136; 0.392]
	Rifafour (N=10)	10	0.150	[0.072; 0.230]

Note: Treatment Group: J = TMC207, J-Z = TMC207 + Pyrazinamide, J-Pa = TMC207 + PA-824, Pa-Z = PA-824 + Pyrazinamide, Pa-Z-M = PA-824 + Pyrazinamide + Moxifloxacin, Rifafour = Rifafour e-275®. EBA$(t_1 - t_2)$: Early bactericidal activity over Day t_1 to Day t_2; BCI: Bayesian credibility interval. n = Number of patients in each category.

Table B.1: Model 1 – Inferential Statistics for Regression Model Parameters

Parameter	Treatment	n	Mean Estimate	95% BCI
$\boldsymbol{\lambda_{1j}}$	J (N=15)	15	-0.002	[-0.088; 0.084]
	J-Z (N=15)	15	0.066	[-0.047; 0.172]
	J-Pa (N=15)	15	0.105	[0.018; 0.187]
	Pa-Z (N=15)	15	0.179	[0.079; 0.278]
	Pa-Z-M (N=15)	15	0.316	[0.159; 0.470]
	Rifafour (N=10)	10	0.155	[0.019; 0.299]
$\boldsymbol{\beta_{2j}}$	J (N=15)	15	0.083	[0.002; 0.167]
	J-Z (N=15)	15	0.051	[-0.038; 0.140]
	J-Pa (N=15)	15	-0.005	[-0.080; 0.073]
	Pa-Z (N=15)	15	-0.030	[-0.118; 0.059]
	Pa-Z-M (N=15)	15	-0.053	[-0.223; 0.136]
	Rifafour (N=10)	10	-0.004	[-0.128; 0.118]
$\boldsymbol{\beta_{2fj}}$	J (N=15)	15	0.083	[-0.213; 0.384]
	J-Z (N=15)	15	0.050	[-0.290; 0.387]
	J-Pa (N=15)	15	-0.004	[-0.268; 0.267]
	Pa-Z (N=15)	15	-0.030	[-0.345; 0.285]
	Pa-Z-M (N=15)	15	-0.054	[-0.651; 0.563]
	Rifafour (N=10)	10	-0.003	[-0.381; 0.388]

Note: Treatment Group: J = TMC207, J-Z = TMC207 + Pyrazinamide, J-Pa = TMC207 + PA-824, Pa-Z = PA-824 + Pyrazinamide, Pa-Z-M = PA-824 + Pyrazinamide + Moxifloxacin, Rifafour = Rifafour e-275®. EBA($t_1 - t_2$): Early bactericidal activity over Day t_1 to Day t_2; BCI: Bayesian credibility interval. n = Number of patients in each category.

Table B.1: Model 1 – Inferential Statistics for Regression Model Parameters

Parameter	Treatment	n	Mean Estimate	Mean 95% BCI
λ_{2j}	J (N=15)	15	0.164	[0.064; 0.275]
	J-Z (N=15)	15	0.167	[0.064; 0.271]
	J-Pa (N=15)	15	0.095	[0.008; 0.184]
	Pa-Z (N=15)	15	0.119	[0.014; 0.219]
	Pa-Z-M (N=15)	15	0.209	[-0.051; 0.492]
	Rifafour (N=10)	10	0.146	[-0.006; 0.299]
κ_j	J (N=15)	15	7.568	[2.797; 10.810]
	J-Z (N=15)	15	4.718	[2.088; 10.030]
	J-Pa (N=15)	15	7.448	[2.642; 10.810]
	Pa-Z (N=15)	15	7.799	[2.614; 10.880]
	Pa-Z-M (N=15)	15	4.561	[2.083; 9.941]
	Rifafour (N=10)	10	5.446	[2.116; 10.570]
γ_j	J (N=15)	15	1.043	[0.147; 1.953]
	J-Z (N=15)	15	1.096	[0.154; 1.958]
	J-Pa (N=15)	15	1.069	[0.150; 1.955]
	Pa-Z (N=15)	15	1.067	[0.149; 1.954]
	Pa-Z-M (N=15)	15	1.029	[0.143; 1.950]
	Rifafour (N=10)	10	1.069	[0.151; 1.955]

Note: Treatment Group: J = TMC207, J-Z = TMC207 + Pyrazinamide, J-Pa = TMC207 + PA-824, Pa-Z = PA-824 + Pyrazinamide, Pa-Z-M = PA-824 + Pyrazinamide + Moxifloxacin, Rifafour = Rifafour e-275®. EBA($t_1 - t_2$): Early bactericidal activity over Day t_1 to Day t_2; BCI: Bayesian credibility interval. n = Number of patients in each category.

Table B.1: Model 1 – Inferential Statistics for Regression Model Parameters

| | | | Mean | |
Parameter	Treatment	n	Estimate	95% BCI
v_j	J (N=15)	15	4.599	[2.115; 12.170]
	J-Z (N=15)	15	3.607	[2.157; 6.437]
	J-Pa (N=15)	15	44.480	[4.053; 96.890]
	Pa-Z (N=15)	15	18.060	[3.133; 86.110]
	Pa-Z-M (N=15)	15	47.630	[5.766; 97.180]
	Rifafour (N=10)	10	10.210	[2.238; 61.090]

Note: Treatment Group: J = TMC207, J-Z = TMC207 + Pyrazinamide, J-Pa = TMC207 + PA-824, Pa-Z = PA-824 + Pyrazinamide, Pa-Z-M = PA-824 + Pyrazinamide + Moxifloxacin, Rifafour = Rifafour e-275®. EBA($t_1 - t_2$): Early bactericidal activity over Day t_1 to Day t_2; BCI: Bayesian credibility interval. n = Number of patients in each category.
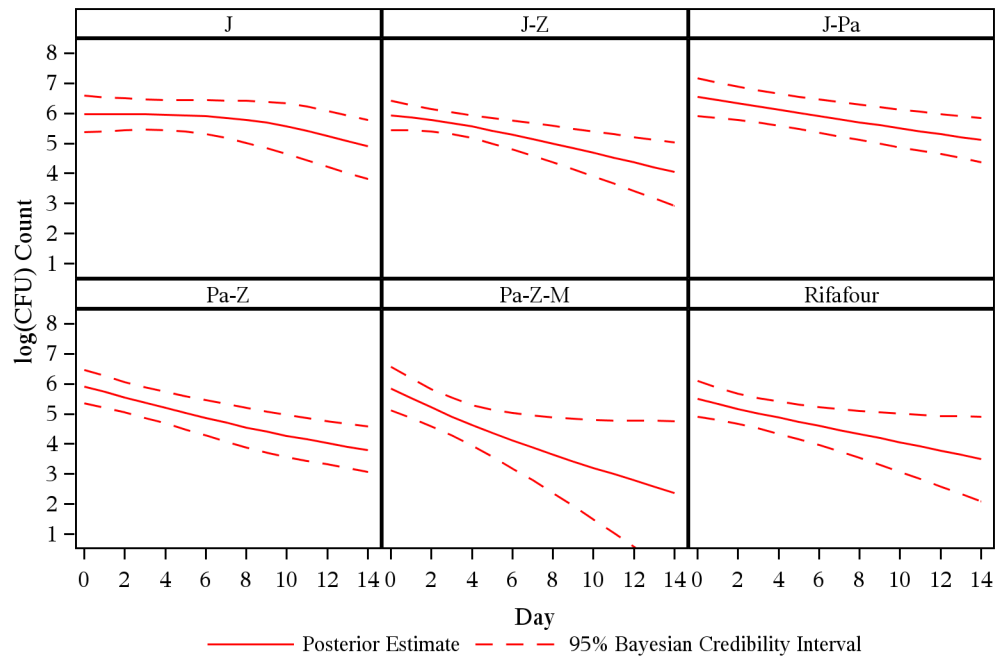
Figure B.1: Model 2 – Posterior Estimates and Corresponding 95% BCIs for Mean log(CFU) Count Over Time
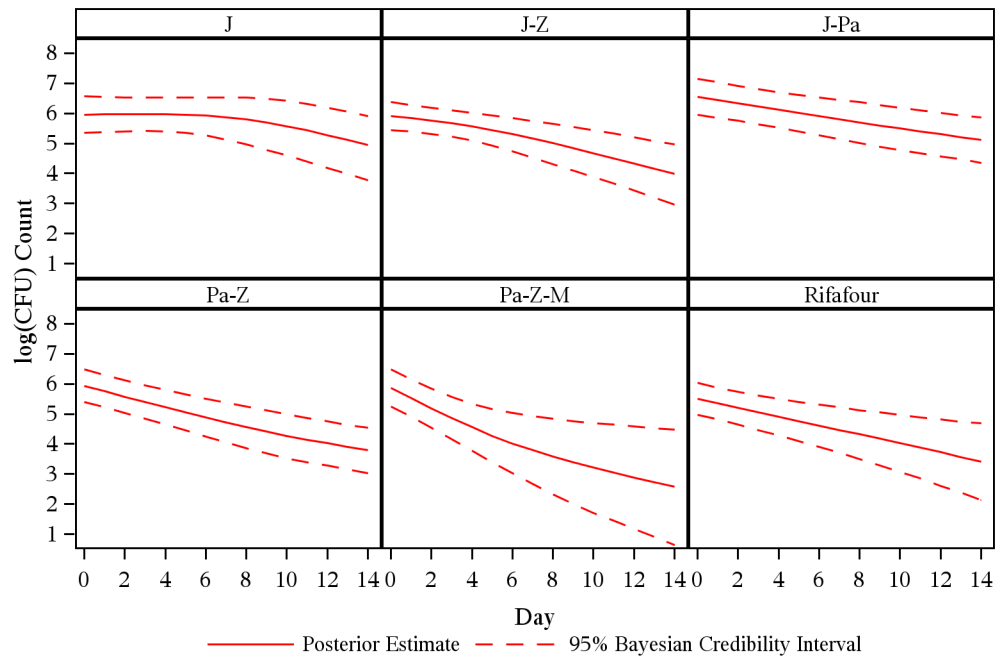
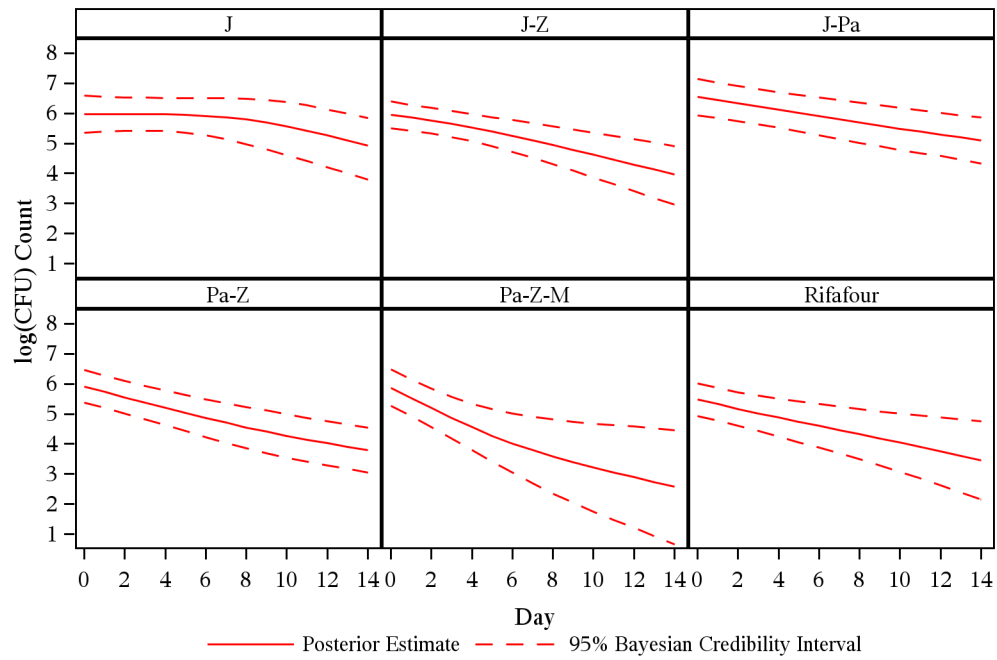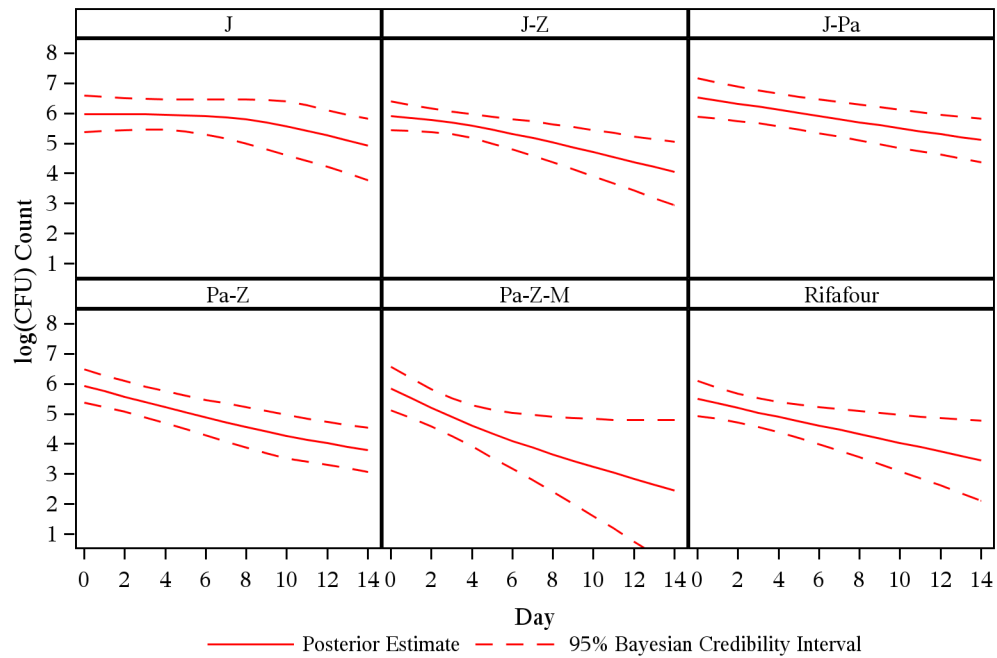**Figure B.2:** Model 3 – Posterior Estimates and Corresponding 95% BCIs for Mean log(CFU) Count Over Time

Figure B.3: Model 4 – Posterior Estimates and Corresponding 95% BCIs for Mean log(CFU) Count Over Time

Figure B.4: Model 5 – Posterior Estimates and Corresponding 95% BCIs for Mean log(CFU) Count Over Time

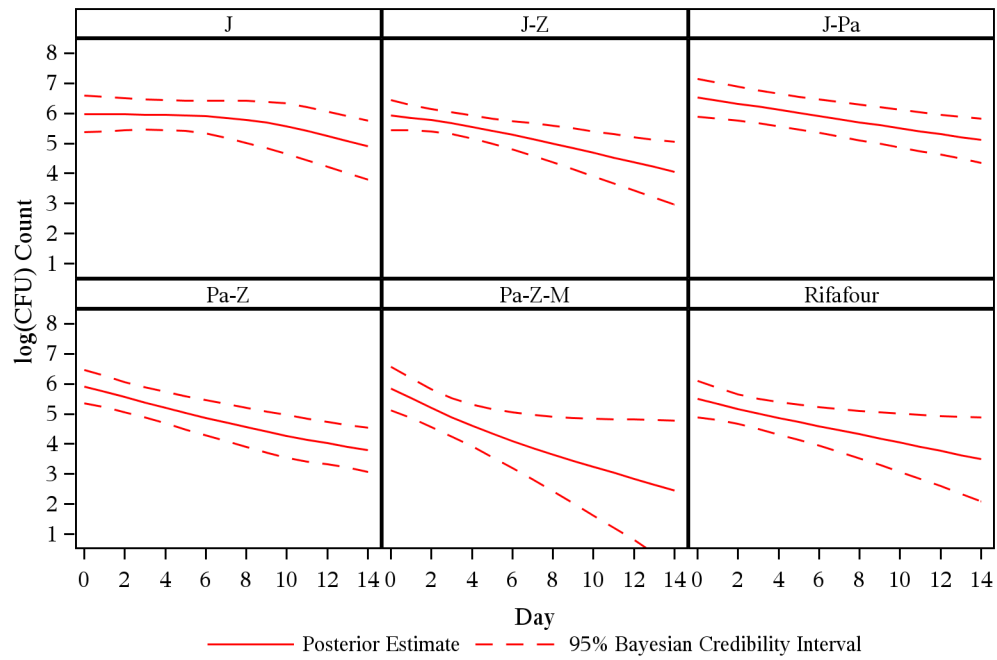**Figure B.5:** Model 6 – Posterior Estimates and Corresponding 95% BCIs for Mean log(CFU) Count Over Time

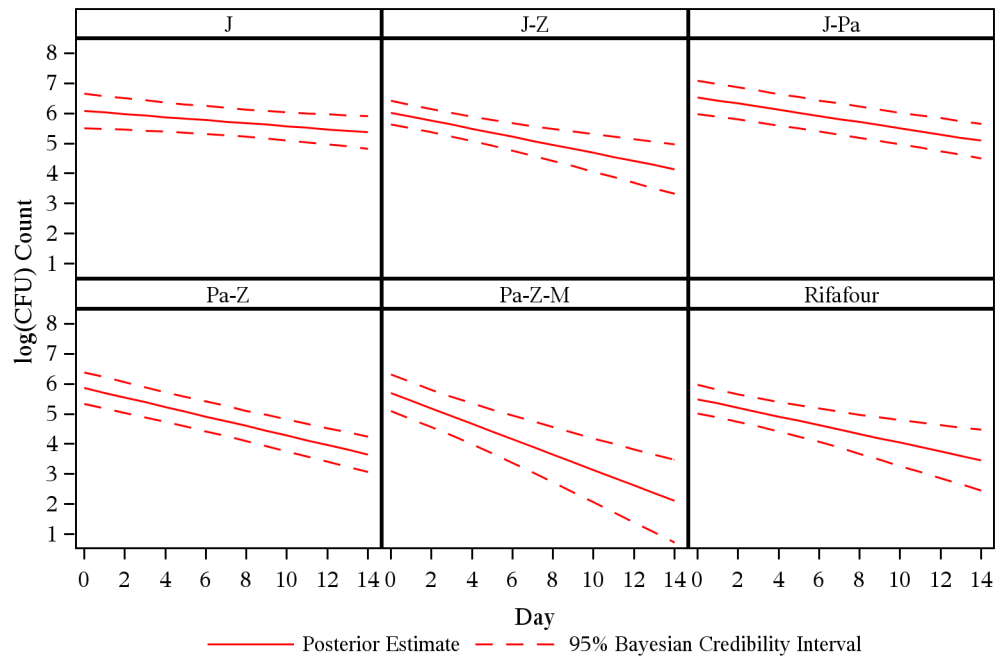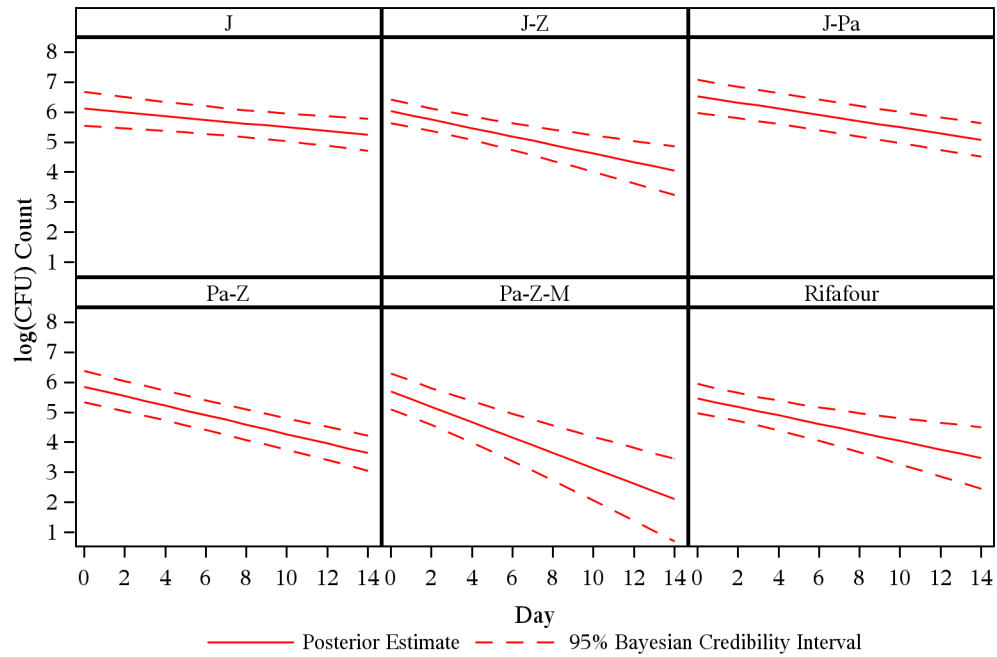**Figure B.6:** Model 7 – Posterior Estimates and Corresponding 95% BCIs for Mean log(CFU) Count Over Time

Figure B.7: Model 8 – Posterior Estimates and Corresponding 95% BCIs for Mean log(CFU) Count Over Time

C MODEL 1: OPENBUGS CODE

```
model {

for (i in 1:1334) {

LOGCFU[i] ~ dt(MEAN[i], INVSIGSQ[KVTRTN[i]], v[KVTRTN[i]])

LOGCFUC[i] ~ dunif(0, 1)

X[i] <- density(LOGCFU[i], DATA[i])

}

for (i in 1335:1360) {

LOGCFUC[i] ~ dt(MEAN[i], INVSIGSQ[KVTRTN[i]], v[KVTRTN[i]])C(, C[i])

X[i] <- cumulative(LOGCFUC[i], C[i])

}

for (i in 1:1360) {

MEAN[i] <- SMU[NSUBJID[i], 1] - SMU[NSUBJID[i], 2]*TIME[i] - SMU[NSUBJID[i], 3]*SGAMMA[NSUBJID[i]]*
          log((exp((TIME[i] - SKAPPA[NSUBJID[i]])/SGAMMA[NSUBJID[i]]) + exp(-(TIME[i] - SKAPPA[NSUBJID[i]])/
          SGAMMA[NSUBJID[i]]))/(exp((SKAPPA[NSUBJID[i]])/SGAMMA[NSUBJID[i]]) + exp(-(SKAPPA[NSUBJID[i]])/
          SGAMMA[NSUBJID[i]])))

PPO[i] <- X[i]

}

for (i in 1:85) {

SALPHA[i] <- SMU[i, 1]

SBETA1[i] <- SMU[i, 2]

SBETA2[i] <- SMU[i, 3]

SLAMBDA1[i] <- SBETA1[i] - SBETA2[i]

SLAMBDA2[i] <- SBETA1[i] + SBETA2[i]

SMU[i, 1:3] ~ dmnorm(MMU[OKVTRTN[1 + 16*(i - 1)], 1:3], MOMGINV[OKVTRTN[1 + 16*(i - 1)], 1:3, 1:3])

SKAPPA[i] ~ dnorm(MKAPPA[OKVTRTN[1 + 16*(i - 1)]], KAPINVSQ[OKVTRTN[1 + 16*(i - 1)]])T(2, 11)

SGAMMA[i] ~ dnorm(MGAMMA[OKVTRTN[1 + 16*(i - 1)]], GAMINVSQ[OKVTRTN[1 + 16*(i - 1)]])T(0.1, 2)

SEBAO02[i] <- SBETA1[i] + SBETA2[i]*SGAMMA[i]*(log(exp((2 - SKAPPA[i])/SGAMMA[i]) + exp(-(2 - SKAPPA[i])/SGAMMA[i])) -
             log(exp((0 - SKAPPA[i])/SGAMMA[i]) + exp(-(0 - SKAPPA[i])/SGAMMA[i])))/2

SEBA007[i] <- SBETA1[i] + SBETA2[i]*SGAMMA[i]*(log(exp((7 - SKAPPA[i])/SGAMMA[i]) + exp(-(7 - SKAPPA[i])/SGAMMA[i])) -
             log(exp((0 - SKAPPA[i])/SGAMMA[i]) + exp(-(0 - SKAPPA[i])/SGAMMA[i])))/7

SEBA014[i] <- SBETA1[i] + SBETA2[i]*SGAMMA[i]*(log(exp((14 - SKAPPA[i])/SGAMMA[i]) + exp(-(14 - SKAPPA[i])/SGAMMA[i])) -
             log(exp((0 - SKAPPA[i])/SGAMMA[i]) + exp(-(0 - SKAPPA[i])/SGAMMA[i])))/14

SEBA214[i] <- SBETA1[i] + SBETA2[i]*SGAMMA[i]*(log(exp((14 - SKAPPA[i])/SGAMMA[i]) + exp(-(14 - SKAPPA[i])/SGAMMA[i])) -
             log(exp((2 - SKAPPA[i])/SGAMMA[i]) + exp(-(2 - SKAPPA[i])/SGAMMA[i])))/12

SEBA714[i] <- SBETA1[i] + SBETA2[i]*SGAMMA[i]*(log(exp((14 - SKAPPA[i])/SGAMMA[i]) + exp(-(14 - SKAPPA[i])/SGAMMA[i])) -
```

```
        log(exp((7 - SKAPPA[i])/SGAMMA[i]) + exp((-(7 - SKAPPA[i])/SGAMMA[i])))/7
}

for (i in 1:6) {

V[i] ~ dunif(2, 100)

MALPHA[i] <- MMU[i, 1]
MBETA1[i] <- MMU[i, 2]
MBETA2[i] <- MMU[i, 3]
MLAMBDA1[i] <- MBETA1[i] - MBETA2[i]
MLAMBDA2[i] <- MBETA1[i] + MBETA2[i]
MMU[i, 1:3] ~ dmnorm(D[1:3], IDENX[1:3, 1:3])
MKAPPA[i] ~ dunif(2, 11)
MGAMMA[i] ~ dunif(0.1, 2)
INVSIGSQ[i] ~ dgamma(0.0001, 0.0001)
SIGSQ[i] <- 1/INVSIGSQ[i]
KAPSIGSQ[i] ~ dunif(0.01, 30)
KAPINVSQ[i] <- 1/KAPSIGSQ[i]
GAMSIGSQ[i] ~ dunif(0.01, 5)
GAMINVSQ[i] <- 1/GAMSIGSQ[i]
SMUTILDA[i, 1:3] ~ dmnorm(MMU[i, 1:3], MOMGINV[i, 1:3, 1:3])
SALPHATILDA[i] <- SMUTILDA[i, 1]
SBETA1TILDA[i] <- SMUTILDA[i, 2]
SBETA2TILDA[i] <- SMUTILDA[i, 3]
MEBA002[i] <- MBETA1[i] + MBETA2[i]*MGAMMA[i]*(log(exp((2 - MKAPPA[i])/MGAMMA[i]) + exp(-(2 - MKAPPA[i])/MGAMMA[i]))) -
              log(exp((0 - MKAPPA[i])/MGAMMA[i]) + exp(-(0 - MKAPPA[i])/MGAMMA[i])))/2
MEBA007[i] <- MBETA1[i] + MBETA2[i]*MGAMMA[i]*(log(exp((7 - MKAPPA[i])/MGAMMA[i]) + exp(-(7 - MKAPPA[i])/MGAMMA[i])) -
              log(exp((0 - MKAPPA[i])/MGAMMA[i]) + exp(-(0 - MKAPPA[i])/MGAMMA[i])))/7
MEBA014[i] <- MBETA1[i] + MBETA2[i]*MGAMMA[i]*(log(exp((14 - MKAPPA[i])/MGAMMA[i]) + exp(-(14 - MKAPPA[i])/MGAMMA[i])) -
              log(exp((0 - MKAPPA[i])/MGAMMA[i]) + exp(-(0 - MKAPPA[i])/MGAMMA[i])))/14
MEBA214[i] <- MBETA1[i] + MBETA2[i]*MGAMMA[i]*(log(exp((14 - MKAPPA[i])/MGAMMA[i]) + exp(-(14 - MKAPPA[i])/MGAMMA[i])) -
              log(exp((2 - MKAPPA[i])/MGAMMA[i]) + exp(-(2 - MKAPPA[i])/MGAMMA[i])))/12
MEBA714[i] <- MBETA1[i] + MBETA2[i]*MGAMMA[i]*(log(exp((14 - MKAPPA[i])/MGAMMA[i]) + exp(-(14 - MKAPPA[i])/MGAMMA[i])) -
              log(exp((7 - MKAPPA[i])/MGAMMA[i]) + exp(-(7 - MKAPPA[i])/MGAMMA[i])))/7

for (j in 0:14) {
  MPLOT[i, j + 1] <- MMU[i, 1] - MMU[i, 2] - MMU[i, 3]*MGAMMA[i]*log((exp((j - MKAPPA[i])/MGAMMA[i]) + exp(-(j - MKAPPA[i])/MGAMMA[i]))/
                     (exp((MKAPPA[i])/MGAMMA[i]) + exp(-(MKAPPA[i])/MGAMMA[i])))
}

}

for (i in 1:6) {
```

```
MEBAD002[i] <- MEBA002[i] - MEBA002[6]
MEBAD007[i] <- MEBA007[i] - MEBA007[6]
MEBAD014[i] <- MEBA014[i] - MEBA014[6]
MEBAD214[i] <- MEBA214[i] - MEBA214[6]
MEBAD714[i] <- MEBA714[i] - MEBA714[6]
}
D[1] <- 0
D[2] <- 0
D[3] <- 0
IDENX[1, 1] <- 0.0001
IDENX[1, 2] <- 0
IDENX[1, 3] <- 0
IDENX[2, 1] <- IDENX[1, 2]
IDENX[2, 2] <- 0.0001
IDENX[2, 3] <- 0
IDENX[3, 1] <- IDENX[1, 3]
IDENX[3, 2] <- IDENX[2, 3]
IDENX[3, 3] <- 0.0001
for (i in 1:6) {
    MOMGINV[i, 1:3, 1:3] ~ dwish(IDEN[i, 1:3, 1:3], 3)
    MOMEGA[i, 1:3, 1:3] <- inverse(MOMGINV[i, 1:3, 1:3])
    ALPSIGSQ[i] <- MOMEGA[i, 1, 1]
    BT1SIGSQ[i] <- MOMEGA[i, 2, 2]
    BT2SIGSQ[i] <- MOMEGA[i, 3, 3]
    ALPBT1SIGSQ[i] <- MOMEGA[i, 1, 2]
    ALPBT2SIGSQ[i] <- MOMEGA[i, 1, 3]
    BT1BT2SIGSQ[i] <- MOMEGA[i, 2, 3]
}
IDEN[1, 1, 1] <- 1.9845083337
IDEN[1, 1, 2] <- 0.1424260547
IDEN[1, 1, 3] <- -0.255846266
IDEN[1, 2, 1] <- IDEN[1, 1, 2]
IDEN[1, 2, 2] <- 0.0216738387
IDEN[1, 2, 3] <- -0.001714245
IDEN[1, 3, 1] <- IDEN[1, 1, 3]
IDEN[1, 3, 2] <- IDEN[1, 2, 3]
IDEN[1, 3, 3] <- 0.1007109971
IDEN[2, 1, 1] <- 3.2813361357
```

```
IDEN[2, 1, 2]    <-   0.23404549
IDEN[2, 1, 3]    <-  -0.42176524
IDEN[2, 2, 1]    <-   IDEN[2, 1, 2]
IDEN[2, 2, 2]    <-   0.0360726551
IDEN[2, 2, 3]    <-  -0.001910575
IDEN[2, 3, 1]    <-   IDEN[2, 1, 3]
IDEN[2, 3, 2]    <-   IDEN[2, 2, 3]
IDEN[2, 3, 3]    <-   0.161357375
IDEN[3, 1, 1]    <-   1.9485281239
IDEN[3, 1, 2]    <-   0.1391632094
IDEN[3, 1, 3]    <-  -0.247040917
IDEN[3, 2, 1]    <-   IDEN[3, 1, 2]
IDEN[3, 2, 2]    <-   0.0213492645
IDEN[3, 2, 3]    <-  -0.000889594
IDEN[3, 3, 1]    <-   IDEN[3, 1, 3]
IDEN[3, 3, 2]    <-   IDEN[3, 2, 3]
IDEN[3, 3, 3]    <-   0.0942288949
IDEN[4, 1, 1]    <-   2.6021610817
IDEN[4, 1, 2]    <-   0.1865544366
IDEN[4, 1, 3]    <-  -0.337417074
IDEN[4, 2, 1]    <-   IDEN[4, 1, 2]
IDEN[4, 2, 2]    <-   0.0291376766
IDEN[4, 2, 3]    <-  -0.001787706
IDEN[4, 3, 1]    <-   IDEN[4, 1, 3]
IDEN[4, 3, 2]    <-   IDEN[4, 2, 3]
IDEN[4, 3, 3]    <-   0.1312407105
IDEN[5, 1, 1]    <-   7.9831351344
IDEN[5, 1, 2]    <-   0.5673254998
IDEN[5, 1, 3]    <-  -1.050960876
IDEN[5, 2, 1]    <-   IDEN[5, 1, 2]
IDEN[5, 2, 2]    <-   0.0944775205
IDEN[5, 2, 3]    <-   0.0019173075
IDEN[5, 3, 1]    <-   IDEN[5, 1, 3]
IDEN[5, 3, 2]    <-   IDEN[5, 2, 3]
IDEN[5, 3, 3]    <-   0.4189948514
IDEN[6, 1, 1]    <-   3.2598540528
IDEN[6, 1, 2]    <-   0.2344166633
IDEN[6, 1, 3]    <-  -0.41072744
```

```
IDEN[6, 2, 1]  <-  IDEN[6, 1, 2]
IDEN[6, 2, 2]  <-  0.0354100862
IDEN[6, 2, 3]  <-  -0.002830085
IDEN[6, 3, 1]  <-  IDEN[6, 1, 3]
IDEN[6, 3, 2]  <-  IDEN[6, 2, 3]
IDEN[6, 3, 3]  <-  0.1559887863
```

}